

RESEARCH ARTICLE

Open Access

# Ancient gene transfer from algae to animals: Mechanisms and evolutionary significance

Ting Ni<sup>1,2†</sup>, Jipei Yue<sup>3†</sup>, Guiling Sun<sup>3</sup>, Yong Zou<sup>1,2</sup>, Jianfan Wen<sup>1\*</sup> and Jinling Huang<sup>3\*</sup>

## Abstract

**Background:** Horizontal gene transfer (HGT) is traditionally considered to be rare in multicellular eukaryotes such as animals. Recently, many genes of miscellaneous algal origins were discovered in choanoflagellates. Considering that choanoflagellates are the existing closest relatives of animals, we speculated that ancient HGT might have occurred in the unicellular ancestor of animals and affected the long-term evolution of animals.

**Results:** Through genome screening, phylogenetic and domain analyses, we identified 14 gene families, including 92 genes, in the tunicate *Ciona intestinalis* that are likely derived from miscellaneous photosynthetic eukaryotes. Almost all of these gene families are distributed in diverse animals, suggesting that they were mostly acquired by the common ancestor of animals. Their miscellaneous origins also suggest that these genes are not derived from a particular algal endosymbiont. In addition, most genes identified in our analyses are functionally related to molecule transport, cellular regulation and methylation signaling, suggesting that the acquisition of these genes might have facilitated the intercellular communication in the ancestral animal.

**Conclusions:** Our findings provide additional evidence that algal genes in aplastidic eukaryotes are not exclusively derived from historical plastids and thus important for interpreting the evolution of eukaryotic photosynthesis. Most importantly, our data represent the first evidence that more anciently acquired genes might exist in animals and that ancient HGT events have played an important role in animal evolution.

**Keywords:** Gene transfer, Endosymbiosis, Plastids, Animal evolution

## Background

Gene transfer, including those occurred horizontally between distinct species (horizontal gene transfer, HGT) or those intracellularly from organelles (mitochondria or plastids) to the nucleus (endosymbiotic gene transfer, EGT), has now been widely recognized as an important force in organismal and genome evolution [1-3]. In particular, HGT may rapidly spread evolutionary success across lineages and allow recipient organisms to access new niches or other resources. Although numerous cases of HGT have been documented in prokaryotes and unicellular eukaryotes, a common belief is that HGT is rare in multicellular eukaryotes such as animals and plants [4,5]. Such a belief, however, largely ignores the dynamic nature of HGT and the more ancient HGT

events to the unicellular ancestors of multicellular eukaryotes [6].

Gene transfer, although usually disruptive in phylogenetic reconstruction, may also serve as molecular fossils or footprints for historical evolutionary events [7,8]. An intriguing case in this respect is the distribution of algal/cyanobacterial genes in eukaryotes. Because eukaryotic photosynthesis is derived from a primary endosymbiosis with a cyanobacterium as well as secondary and tertiary endosymbioses with miscellaneous algae [9-12], these genes are frequently interpreted as relicts of earlier cyanobacterial/algal endosymbionts and, therefore, evidence of plastid losses in some aplastidic eukaryotes [13-15]. As eukaryotic photosynthesis spans multiple major lineages whose relationships critically rely on the plastid existence in them [16-18], an accurate understanding of the distribution and origin of algal genes in eukaryotes is essential.

Several recent studies indicated that algal genes also exist in organisms that are generally considered to be plastid-lacking throughout their evolution [15,19-21]. In

\* Correspondence: wenjfi@mail.kiz.ac.cn; huangj@ecu.edu

†Equal contributors

<sup>1</sup>State Key Laboratory of Genetic Resources and Evolution, Kunming Institute of Zoology, Chinese Academy of Sciences, Kunming, Yunnan 650223, China  
Full list of author information is available at the end of the article

particular, over 100 genes of algal origins were found in the choanoflagellate *Monosiga brevicollis* [19,21]. Because of the widespread distribution of algae and the phagotrophic nature of choanoflagellates, it was suggested that algal genes in *Monosiga* might have been acquired from food sources [21]. However, because choanoflagellates are the closest unicellular relatives of animals [22], it is unclear whether such acquisition of algal genes represents species-specific events or might have occurred also in the unicellular ancestor of animals and been inherited by diverse extant animals.

To address the above question, we performed phylogenomic analyses of the tunicate *Ciona intestinalis* to search for algal genes, in particular those anciently acquired by the ancestral animal. We choose *C. intestinalis* as a query to detect algal genes in animals because of its relatively small genome size and frequent physical contact with phytoplanktons. We here report 14 gene families of likely algal origin in *C. intestinalis* and other animals. We also further discuss the potential mechanisms of algal gene acquisition in animals and the importance of HGT in animal evolution.

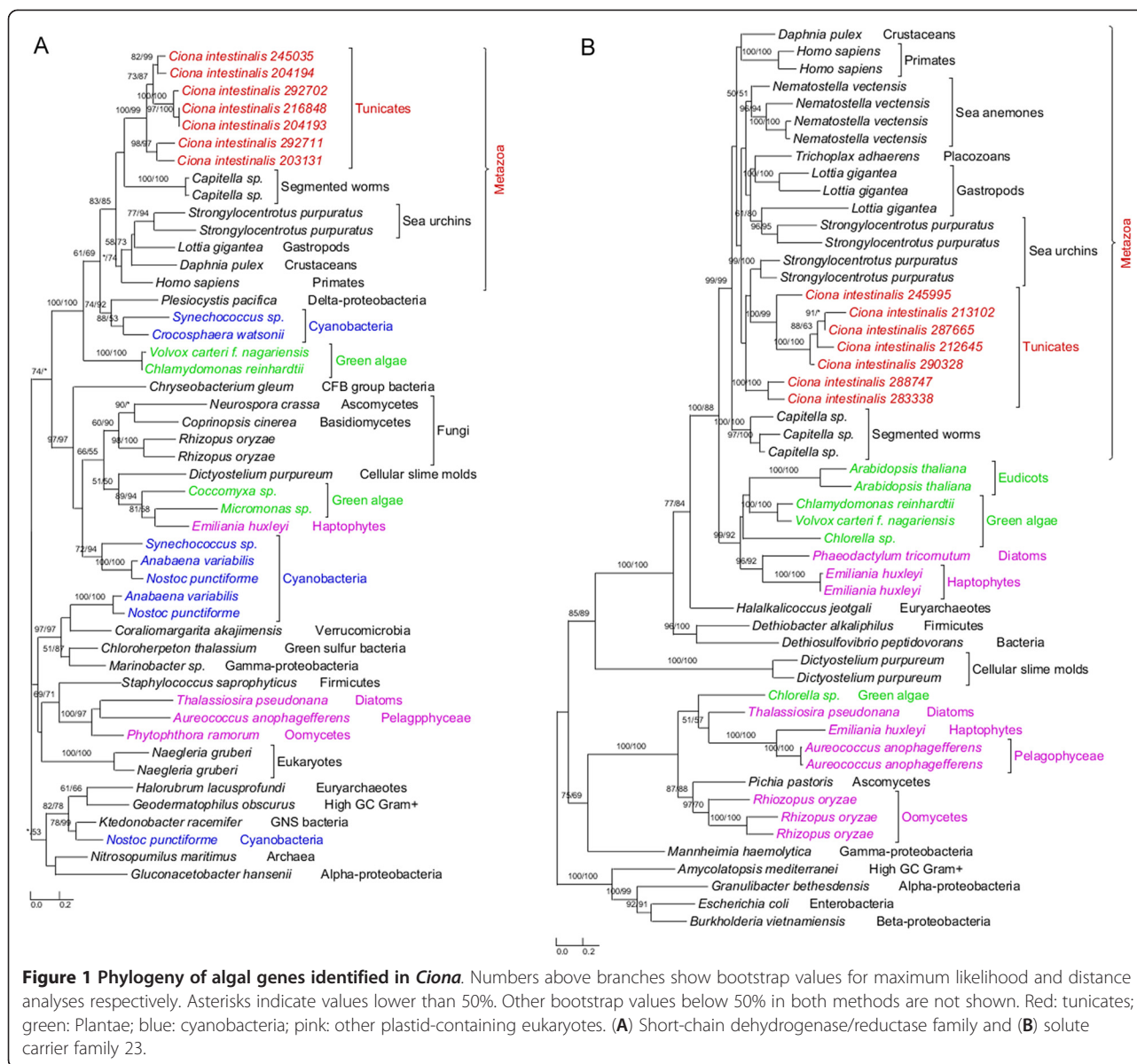
## Results and discussion

### Algae-related genes identified in *C. intestinalis* and other animals

The annotated genome of *C. intestinalis* consists of 14,002 protein-coding genes and genome screening using *AlienG* [23] identified 169 of them as candidates of algal origin. We performed detailed analyses of taxonomic distribution, gene structure and molecular phylogeny for each of the identified candidate genes. One of these genes (GenBank accession number: XP\_002127943) has identifiable homologs only in animals and plastid-containing eukaryotes. Another identified gene (GenBank accession number: XP\_002122576) is restricted to animals, plastid-containing eukaryotes and ciliates, which also contain algal genes because of their possible historical plastids or phagotrophic lifestyle [14]. For additional 90 genes, although their molecular phylogenies often lack sufficient resolution in some branches, algal and animal sequences form monophyletic groups with over 70% bootstrap support (Figures 1, 2 and 3). Therefore, 92 of the 169 genes are considered to be algae-related. Additional evidence from Pfam domain analyses [24] also indicates animal/algae affinity. For instance, three genes encoding potassium/sodium hyperpolarization-activated cyclic nucleotide-gated channel 2 contain three identical tandem domains only in animals and algae (Figure 3). These 92 genes can be further classified into 14 gene families or subfamilies, including those encoding solute carriers, short-chain dehydrogenase/reductase, and UDP

galactopyranose mutase etc (Table 1), based on their sequence relatedness (30-80% protein sequence identities and similar tree topologies) and GO annotation information (Additional file 1). Except the genes encoding UDP galactopyranose mutase, all other algae-related genes are present in multiple groups of animals (Figures 1, 2 and 3; Additional file 2).

Each of the 14 gene families identified in our analyses shows a complex evolutionary history that differs from perceived organismal relationships in three domains of life. In many cases, multiple eukaryotic sequences are sporadically clustered within bacterial homologs. Such a complex evolutionary history for individual gene (or gene family) is somewhat expected, considering frequent HGT events both within domains (e.g., bacteria and archaea) [25,26] and between domains (e.g., bacteria or archaea to unicellular eukaryotes) [27-29], EGT following primary and higher level endosymbioses [3,30], and differential genes losses or replacements [31,32] etc. The dynamic occurrence of these factors over evolutionary time may further compound their effects. Therefore, aside from potential phylogenetic artifacts (see next paragraph), the monophyletic relationship between algal and animal sequences observed in this study can largely be explained by two alternative scenarios: (i) gene losses in most protists and fungi and (ii) gene acquisition by animals. For example, genes encoding UDP galactopyranose mutase are restricted to *C. intestinalis*, algae, and bacteria (including cyanobacteria). Phylogenetic analyses show that *Ciona* UDP galactopyranose mutase sequences group with homologs from multiple algae (Additional file 2). This may be attributed to either extensive gene losses from other eukaryotes or unique gene acquisition by *Ciona*. The scenario of extensive gene losses from other eukaryotes, though theoretically possible, is not parsimonious [6]. Gene acquisition followed by duplication appears to be a more plausible explanation for the presence of this gene family in *Ciona*. Other phylogenetic trees provide similar topologies, where *Ciona* sequences form monophyletic groups with homologs from algae and other animals (Figures 1, 2 and 3; Additional file 2). For some identified gene families, other lines of evidence also argue strongly against the scenario of differential gene losses in other eukaryotes; such evidence includes 1) the presence of identifiable homologs only in animals and plastid-containing eukaryotes; 2) the monophyly of animal sequences within algal homologs; 3) uniquely shared domain structures between animal and algal sequences e.g. (Figure 3); 4) the availability of a large number of fungal genomes and the absence of some identified genes in fungi. Furthermore, two of the 14 gene families show cyanobacterial/plastidic affiliation (Figures 1A and 2A), suggesting likely algal origin. Several other gene families also encode proteins that are targeted to chloroplasts in



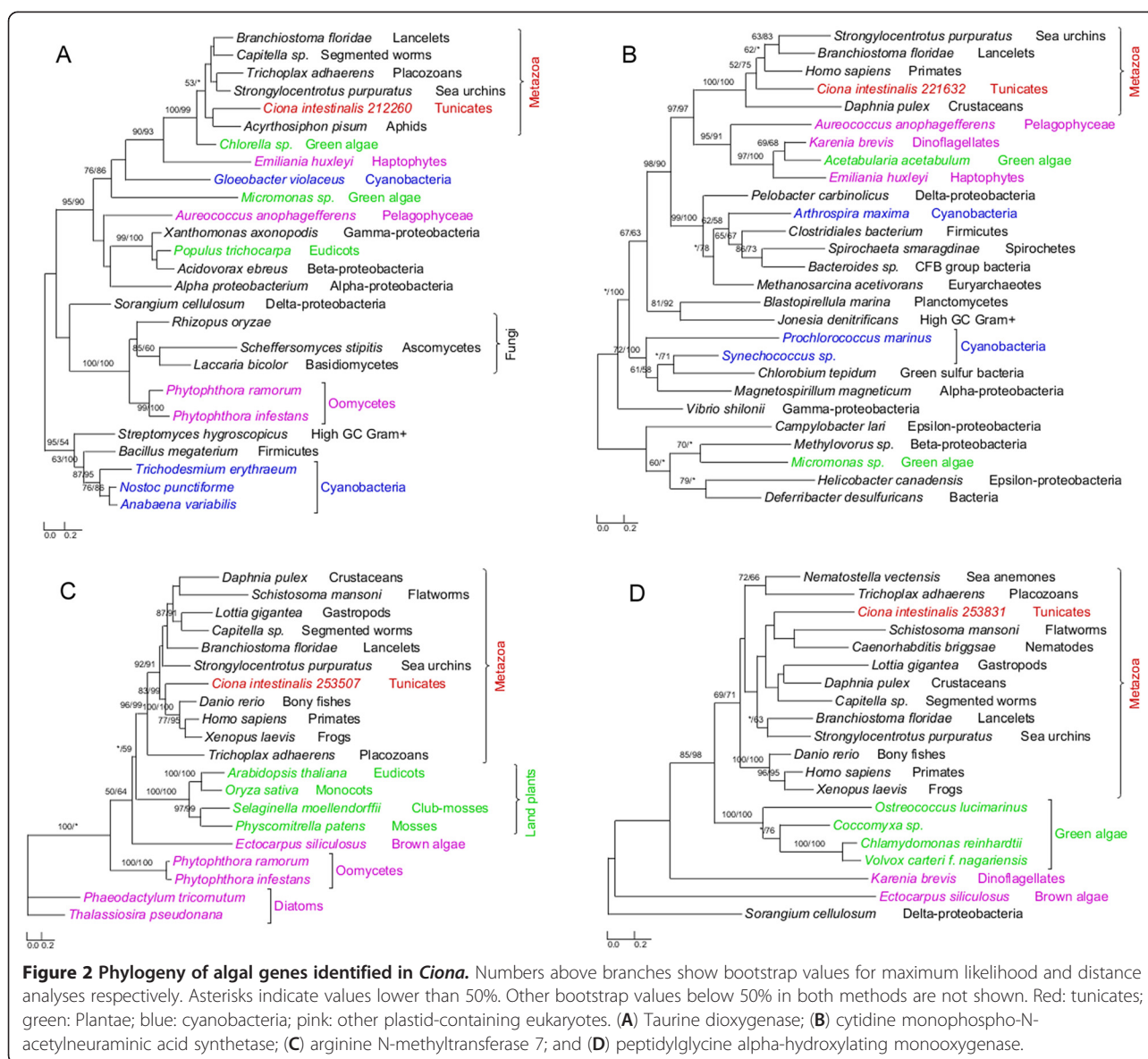
green plants (e.g., *Arabidopsis thaliana*, *Chlamydomonas reinhardtii* and *Micromonas pusilla*) (Additional file 1).

Accurate identification of HGT may be affected by multiple factors. Improper methodology, poor data quality, and insufficient or biased taxonomic sampling are among the factors leading to incorrect gene tree topologies [33]. Especially, long-branch attraction, which may occur when two sequences evolved much faster or slower than others, demands particular attention [34]. In order to reduce the potential complications resulting from these issues, we searched the NCBI non-redundant (*nr*) protein sequence database, ESTs and other sequence data to ensure a broad sequence sampling. We further examined the data quality and performed careful phylogenetic analyses by selecting the optimal substitution matrix for each dataset and using

different computational algorithms. Nevertheless, with improved phylogenetic algorithms and more sequence data from other eukaryotes becoming available, we expect that the tree topologies for the identified genes may vary. It should also be noted that, because the phylogenetic approach tends to underestimate the number of acquired genes and because the *Ciona* genome was the sole query in our analyses, it is likely that other algae-related genes may exist in animals. Therefore, the identified genes in our analyses should not be considered as a definitive list of algae-related genes in animals.

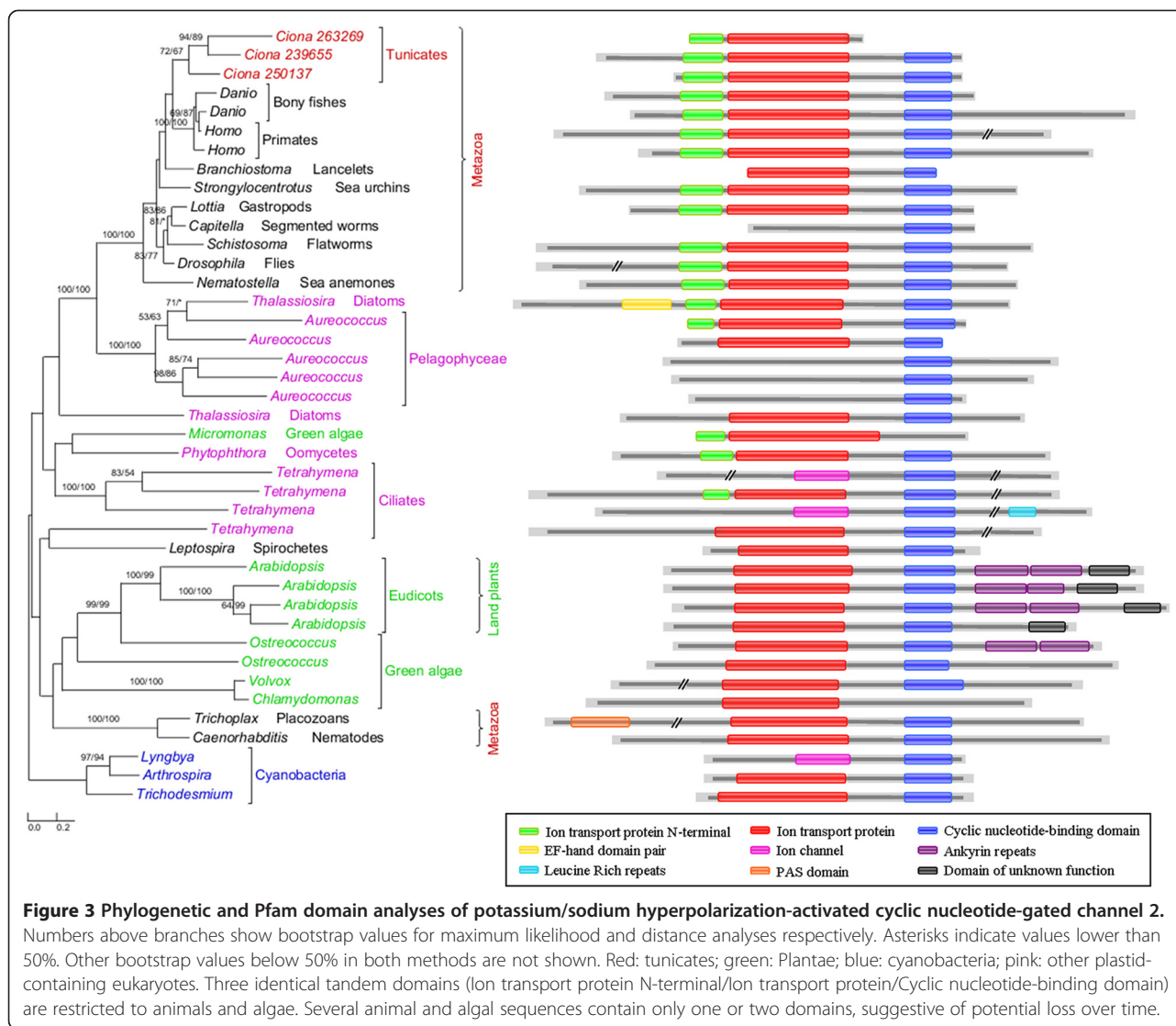
#### Algal genes and the historical distribution of plastids

Given their presence and relatedness in multiple animal groups, the 14 genes families identified in this study, if



indeed of algal origin (algal genes hereafter), most likely were acquired by the ancestral animal. This ancestral animal was probably unicellular, like its extant closest relatives choanoflagellates [22]. The identified gene families might have been acquired through either symbioses or feeding activities. In the first scenario, the ancestral animal acquired algal genes as a result of endosymbioses with algal cells, followed by loss of plastids. The second scenario suggests that algal genes were routed to the nucleus of the ancestral animal after algal cells were phagocytosed. In most symbioses between algae and animals, algal symbionts either form loose extracellular association with animals or are sequestered in vacuoles of host cells [35]. Thus far, neither scenario is known to lead to gene transfers. Corals and salamander *Ambystoma maculatum*, for example, enjoy an endosymbiotic

relationship with the dinoflagellate *Symbiodinium* and the green alga *Oophila amblystomatis*, respectively [36,37]. But neither EGT between corals and dinoflagellates nor between salamanders and green algal endosymbionts has been reported. By far, only the endosymbioses in sea slug *Elysia chlorotica* and cnidarian *Hydra viridis* are reportedly accompanied by transfer of algal genes [35,38-40]. Because such endosymbioses-mediated gene transfer is unknown in other animals, it might be extremely rare. Given the number of algal gene families identified in our study and their miscellaneous donor groups (Table 1; Figures 1, 2 and 3; Additional file 2), it is highly improbable that these gene families are derived from such rare events. On the other hand, because most of the algal gene families identified in animals are derived from microscopic phytoplanktons that are



widely distributed in aquatic environments, it is plausible that the ancestral animal obtained these algal genes via food capture.

The algal gene families identified in animals and their possible acquisition from food sources bear important implications for interpreting the historical distribution of plastids and eukaryotic photosynthesis. The origin of plastids is one of the most important evolutionary events in eukaryotic evolution. However, the historical distribution of plastids and photosynthesis in eukaryotes is still heatedly debated [14,16-18,41,42]. Because of the potential loss of plastids, algal genes in aplastidic eukaryotes have often been interpreted as derived from historical plastids. For example, algal genes in apicomplexan *Cryptosporidium*, heterokont *Phytophthora*, and ciliates have been cited as evidence for plastid losses in these taxa [14,33,43]. Similar arguments were also made for kinetoplastids based on the presence of “plant-like”

genes [44], though whether these genes are indeed of plant origin is questionable [45,46]. Historical existence of plastids was also suggested for some protozoan species such as heterolobosean *Naegleria* [15,47]. Although endosymbiosis indeed appears to be a reasonable explanation for the observed data in some cases, recent findings of widespread HGT events in eukaryotes raise serious questions about whether all algal genes are derived from historical algal endosymbionts or plastids.

Many studies show that HGT from bacteria is frequent in unicellular eukaryotes [28,29,48]. There is also convincing evidence for HGT between eukaryotes [4,49-51]. In particular, recent studies identified over 100 algal genes in the choanoflagellate *Monosiga* [19,21]. The number of algal genes in *Monosiga* is considerably greater than those in many other aplastidic eukaryotes (e.g., ciliates, *Cryptosporidium*, *Naegleria* etc) that have been suggested to contain historical plastids. Although

**Table 1 Putative algal genes identified in *Ciona intestinalis***

Gene family <sup>a</sup>	Putative donor	Putative function
Short-chain dehydrogenase/reductase family (7)	Green algae/ cyanobacteria	Cellular differentiation and signaling
Solute carrier family 23 (7)	Algae/land plants	Na <sup>+</sup> -dependent vitamin C transport
Taurine dioxygenase (1)	Green algae	Cell redox homeostasis
Cytidine monophospho-N-acetylneuraminic acid synthetase (1)	Algae	Signaling regulation
Arginine N-methyltransferase 7 (1)	Land plants/ stramenopiles	Protein methylation
Peptidylglycine alpha-hydroxylating monooxygenase (1)	Green algae	Hormone biosynthesis
Aminoglycoside phosphotransferase (1)	Algae	Protein amino acid phosphorylation
Solute carrier family 22 (27)	Algae/land plants	Ion transport
Solute carrier family 34 (6)	Diatoms	Phosphate ion transport
Potassium/sodium hyperpolarization-activated cyclic nucleotide-gated channel 2 (3)	Stramenopiles	Potassium ion transport
UDP galactopyranose mutase (2)*	Algae	Uridine diphosphogalactofuranose biosynthesis
Biphenyl/valacyclovir hydrolase (3)	Dinoflagellates/haptophytes	Aromatic compound metabolism
Solute carrier family 6 (31)	Algae	Neurotransmitter transport
Alkylation repair homolog 5 (1)	Algae/land plants	Alkylation repair

<sup>a</sup>Numbers in the brackets show paralogous gene copies within each family. \*The gene transfer event happened only between algae and *C. intestinalis*. All other gene transfers happened between plastid-containing organisms and the ancestral animal.

the possibility that algal genes in *Monosiga* are derived from an obsolete algal endosymbiont or plastid cannot be confidently excluded, the phagotrophic lifestyle of *Monosiga* points to the possibility that these algal genes are derived from food sources. Considering that genes from other prokaryotes can be horizontally acquired by eukaryotes, it should not be surprising to see cyanobacterial (or plastidic) genes in aplastidic eukaryotes. Likewise, it should not be expected that algal genes are immune to HGT, considering that HGT events do occur between other eukaryotes. This may be particularly true for phagotrophic organisms feeding on miscellaneous microbes including unicellular algae. In such cases, algal genes in aplastidic eukaryotes may not necessarily represent relics of historical plastids. These genes should be treated equally as other HGT-derived genes when they are used to infer historical endosymbionts [41], unless other independent evidence (e.g. plastid existence in closely related taxa and compatible phylogenetic signals from multiple genes) is available. Our finding of algal genes in animals provides additional evidence about the distribution of algal genes in aplastidic eukaryotes. Additionally it is consistent with the earlier suggestion that algal genes are expected in phagotrophic eukaryotes or descendants of phagotrophic ancestors [21].

#### Evolutionary significance of algal genes in animals

Several cases of HGT between bacterial endosymbionts and their animal hosts have been documented [52-55]. Moreover, HGT from other free-living organisms has also been reported in cnidarians [39,56,57], bdelloid

rotifers [58], insects [59], and nematodes [60-62] and other animals [63]. The acquired genes are often linked to novel phenotypes and abilities in recipient organisms, such as improved preying capability in cnidarians [56] and reduced susceptibility to predation in aphids [59]. These HGT cases predominantly occurred more recently in taxa of lower taxonomic ranks and, therefore, do not affect the evolution of entire animal lineage.

Despite the increasing number of HGT-derived genes reported in animals, a common belief is that HGT in animals is rare and its role is limited. This belief largely stems from the fact that the acquired genes need to overcome the germline barrier in order to be transmitted to next generations [4,5]. However, such a germline barrier should not exist in the unicellular ancestor of animals and it is conceivable that the ancestral animal, like its closest relatives choanoflagellates or many other unicellular eukaryotes [29,64], was subject to more frequent HGT. In our study, 13 of the 14 gene families identified in *Ciona* are most likely derived from ancient HGT events. These anciently acquired genes have been vertically inherited, duplicated and retained in diverse groups of extant animals, and thus have contributed to the long-term evolution of animals. Such frequent duplication following gene acquisition has also been observed in other studies [62,65-68] and, in many cases, associated with positive selection in recipient organisms [66,67].

Compared to *Monosiga*, the algal gene families identified in animals are considerably fewer and differ distinctly in their functions. Given the limited number of

choanoflagellate genomes available, it is unclear what percentage of the identified algal gene families in *Monosiga* were anciently acquired. However, the lower number of anciently acquired genes in animals is consistent with the suggestion that most acquired genes will eventually be deleted from the recipient genome over time [69]. While the algal genes in *Monosiga* are predominantly involved in carbohydrate and amino acid metabolism [21], five of the 14 gene families, which account for 80% (74/92) of algal genes identified in *Ciona*, are functionally related to molecule transport. The vast majority of these transporter genes (72/74) belong to four solute carrier (SLC) families, including SLC6, SLC22, SLC23 and SLC34 (Table 1), which include transporters of neurotransmitter, organic cation, zwitterion/cation, organic anion, Na<sup>+</sup>-dependent vitamin C transport and phosphate ion, participating in uptake or excretion of numerous important compounds [70-73]. Besides these SLC families, another identified algal gene family is involved in potassium ion transport [74] (Table 1). As these genes encode multiple membrane-bound transporters, substrates such as amino acids, oligopeptides, sugars, inorganic cations and anions, essential metals, biogenic amines, vitamins, nucleosides and ammonia can be transferred across membranes, which in turn accelerates intercellular communication and leads to more efficient cellular metabolism. Several other acquired algal gene families in animals encode proteins that are functionally related to signaling and hormone biosynthesis. For example, the large short-chain dehydrogenase/reductase family (SDR) (Table 1) encodes enzymes involved in cellular differentiation and signaling [75]. Additionally, the biphenyl/valacyclovir hydrolase family is related to aromatic compound metabolism, which may allow the ancestral animal to access diverse digestible food sources.

As all functional information for the algal gene families identified in our analyses is based on investigations of homologs in humans and model organisms, whether these genes have exactly identical functions in other animals remains to be further studied. However, considering the high percent identifies (30%-80%) and close affinities between homologs from humans and other animals (Figures 1, 2, and 3, Additional file 1), it is likely that these genes have similar primary functions. Even if the identified genes might have different functions in other animals, they could provide raw stocks for gene and functional differentiation. Extensive intercellular communication is a major feature distinguishing animals from unicellular eukaryotes, and such communication requires proteins related to molecule transport and signal transduction in multicellular animals [76,77]. The acquisition of genes related to molecular transport and signaling and their subsequent duplication might have

facilitated the gene renovation and multicellular development in animals. Therefore, our findings, although based solely on analyses of algal genes in a relative small genome of *Ciona*, point to an important contribution of HGT to animal evolution.

## Conclusions

Phylogenomic analyses of the tunicate *C. intestinalis* provide evidences for the existence of algal genes in animals. However, the existence of algal genes in animals does not necessarily constitute evidence for historical occurrence of plastids. Almost all algal gene families identified in our analyses were likely transferred to the ancestral animal and duplicated afterward. Most identified algal genes are related to molecule transport and signaling, suggesting their important role in intercellular communication and possibly the origin of multicellularity in animals.

## Methods

### Data sources and genome screening

The annotated genome of the tunicate *C. intestinalis* was downloaded from the Joint Genome Institute (<http://genome.jgi-psf.org/Cioin2/Cioin2.home.html>). Expressed sequence tags (ESTs) of 50 diverse eukaryotes (Additional file 3) were downloaded from the NCBI dbEST database and the Taxonomically Broad EST Database (TBestDB) [78]. Assembly of these ESTs was carried out using CAP3 [79]. The resulting consensus sequences were translated in all six frames using *transeq* of the EMBOSS package [80]. All other sequences used in the analyses were retrieved from the NCBI *nr* protein sequence database.

A comprehensive database was created by including the *nr* database and other available eukaryotic genomes (Additional file 3). Genome screening for candidates of HGT-derived genes was performed using *AlienG* [23]. *AlienG* presumes that sequence similarity is correlated with sequence relatedness and it identifies candidates of acquired genes by comparing query sequence similarity to homologs from potential donors and those from closely related taxa. Genes with identifiable homologs only in potential donors are also identified by *AlienG*. In this study, candidate genes of algal origin in *C. intestinalis* were obtained if they showed significantly higher sequence similarity to algal or cyanobacterial homologs than to those from other eukaryotes; the significant sequence similarity to algal homologs was empirically set to a bit score ratio of over 1.5.

### Phylogenetic analyses

For each gene candidate predicted by *AlienG*, we performed further detailed analyses of taxonomic distribution, gene structure, and molecular phylogeny. Protein

sequences were sampled from representative groups within each domain of life (bacteria, archaea, and eukaryotes) based on the blastp results against *nr* database. To ensure that sequences were sampled as broadly as possible, we also retrieved sequences from other available eukaryotic genomes and EST databases (see above section).

Multiple protein sequence alignments were performed using ClustalX [81,82] under the default settings, followed by manual refinement. Misaligned sequences, gaps and ambiguous sites were removed manually. The alignment data are available upon request. Phylogenetic analyses were performed with a maximum likelihood method using PhyML 3.0 [83] and a distance method using PHYLIP 3.69 [84]. ModelGenerator [85] was used to select the available model of protein substitution and rate heterogeneity that best fit each dataset. Bootstrap analyses with 100 pseudoreplicates were performed in both methods. Programs from the PHYLIP 3.69 package were used to create pseudoreplicate datasets (SEQBOOT), compute distance matrix (PROTDIST), calculate distance trees (NEIGHBOR), and generate the bootstrap consensus tree (CONSENSE). Gene trees were depicted by combining phylogenies from PhyML 3.0 and PHYLIP 3.69 using TreeGraph 2 [86,87].

## Additional files

**Additional file 1: Table S1 and Table S2.** Gene identifiers, chromosomal locations and functions for algal gene families identified in this study.

**Additional file 2: Figure S1-S8.** Molecular phylogenies of algal genes identified in *Ciona intestinalis*.

**Additional file 3: Document.** List of 50 eukaryotes whose ESTs were used and other 14 complete genome sequences were used in this study, in addition to the NCBI *nr* database.

## Competing interests

The authors declare that they have no competing interests.

## Authors' contributions

JFW and JH conceived and designed the study and wrote manuscript. TN performed the analyses and wrote the manuscript. JY participated in the data interpretation and wrote the manuscript. GLS and YZ contributed to the data generation. All authors read and approved the final manuscript.

## Acknowledgements

We thank two anonymous reviewers for their helpful comments and suggestions. This work is supported in part by 973 Program (2007CB815705), the Knowledge Innovation Program (KSCX2-EW-J-23), grant (30830018) from the NSFC, and grant (2008GA002) from Yunnan province to JW and a NSF Assembling the Tree of Life (ATOL) grant (DEB 0830024) to JH.

## Author details

<sup>1</sup>State Key Laboratory of Genetic Resources and Evolution, Kunming Institute of Zoology, Chinese Academy of Sciences, Kunming, Yunnan 650223, China. <sup>2</sup>Graduate School of the Chinese Academy of Sciences, Beijing 100039, China. <sup>3</sup>Department of Biology, East Carolina University, Greenville, NC 27858, USA.

Received: 21 March 2012 Accepted: 1 June 2012  
Published: 12 June 2012

## References

1. Keeling PJ, Palmer JD: Horizontal gene transfer in eukaryotic evolution. *Nat Rev Genet* 2008, **9**(8):605–618.
2. Gogarten JP, Doolittle WF, Lawrence JG: Prokaryotic evolution in light of gene transfer. *Mol Biol Evol* 2002, **19**(12):2226–2238.
3. Martin W, Rujan T, Richly E, Hansen A, Cornelsen S, Lins T, Leister D, Stoebe B, Hasegawa M, Penny D: Evolutionary analysis of Arabidopsis, cyanobacterial, and chloroplast genomes reveals plastid phylogeny and thousands of cyanobacterial genes in the nucleus. *Proc Natl Acad Sci U S A* 2002, **99**(19):12246–12251.
4. Andersson JO: Lateral gene transfer in eukaryotes. *Cell Mol Life Sci* 2005, **62**(11):1182–1197.
5. Kurland CG, Canback B, Berg OG: Horizontal gene transfer: A critical view. *Proc Natl Acad Sci U S A* 2003, **100**(17):9658–9662.
6. Huang JL, Gogarten JP: Ancient horizontal gene transfer can benefit phylogenetic reconstruction. *Trends in Genet* 2006, **22**(7):361–366.
7. Huang JL, Gogarten JP: Did an ancient chlamydial endosymbiosis facilitate the establishment of primary plastids? *Genome Biol* 2007, **8**(6):R99.
8. Moustafa A, Beszteri B, Maier UG, Bowler C, Valentin K, Bhattacharya D: Genomic Footprints of a Cryptic Plastid Endosymbiosis in Diatoms. *Science* 2009, **324**(5935):1724–1726.
9. Delwiche CF: Tracing the thread of plastid diversity through the tapestry of life. *Am Nat* 1999, **154**:S164–S177.
10. Keeling PJ: Diversity and evolutionary history of plastids and their hosts. *Am J Bot* 2004, **91**(10):1481–1493.
11. Bhattacharya D, Yoon HS, Hackett JD: Photosynthetic eukaryotes unite: endosymbiosis connects the dots. *Bioessays* 2004, **26**(1):50–60.
12. Lane CE, Archibald JM: The eukaryotic tree of life: endosymbiosis takes its TOL. *Trends in Ecol & Evol* 2008, **23**(5):268–275.
13. Maruyama S, Misawa K, Iseki M, Watanabe M, Nozaki H: Origins of a cyanobacterial 6-phosphogluconate dehydrogenase in plastid-lacking eukaryotes. *BMC Evol Biol* 2008, **8**:151.
14. Reyes-Prieto A, Moustafa A, Bhattacharya D: Multiple genes of apparent algal origin suggest ciliates may once have been photosynthetic. *Curr Biol* 2008, **18**(13):956–962.
15. Maruyama S, Matsuzaki M, Misawa K, Nozaki H: Cyanobacterial contribution to the genomes of the plastid-lacking protists. *BMC Evol Biol* 2009, **9**:197.
16. Cavalier-Smith T: Principles of protein and lipid targeting in secondary symbiogenesis: Euglenoid, dinoflagellate, and sporozoan plastid origins and the eukaryote family tree. *J Eukaryot Microbiol* 1999, **46**(4):347–366.
17. Nozaki H, Matsuzaki M, Takahara M, Misumi O, Kuroiwa H, Hasegawa M, Shin-i T, Kohara Y, Ogasawara N, Kuroiwa T: The phylogenetic position of red algae revealed by multiple nuclear genes from mitochondria-containing eukaryotes and an alternative hypothesis on the origin of plastids. *J Mol Evol* 2003, **56**(4):485–497.
18. Nozaki H: A new scenario of plastid evolution: plastid primary endosymbiosis before the divergence of the "Plantae," emended. *J Plant Res* 2005, **118**(4):247–255.
19. Nedelcu AM, Miles IH, Fagir AM, Karol K: Adaptive eukaryote-to-eukaryote lateral gene transfer: stress-related genes of algal origin in the closest unicellular relatives of animals. *J Evol Biol* 2008, **21**(6):1852–1860.
20. Nedelcu AM, Blakney AJC, Logue KD: Functional replacement of a primary metabolic pathway via multiple independent eukaryote-to-eukaryote gene transfers and selective retention. *J Evol Biol* 2009, **22**(9):1882–1894.
21. Sun GL, Yang ZF, Ishwar A, Huang JL: Algal Genes in the Closest Relatives of Animals. *Mol Biol Evol* 2010, **27**(12):2879–2889.
22. Lang BF, O'Kelly C, Nerad T, Gray MW, Burger G: The closest unicellular relatives of animals. *Curr Biol* 2002, **12**(20):1773–1778.
23. Tian J, Sun GL, Ding Q, Huang JL, Oruganti S, Xie B: *AlienG: an effective computational tool for phylogenetic identification of horizontally transferred genes*. New Orleans, Louisiana: The third International Conference on Bioinformatics and Computational Biology (BiCoB); 2011.
24. Finn RD, Mistry J, Tate J, Coghill P, Heger A, Pollington JE, Gavin OL, Gunasekaran P, Ceric G, Forslund K, et al: The Pfam protein families database. *Nucleic Acids Res* 2010, **38**:D211–D222.
25. Ochman H, Lawrence JG, Groisman EA: Lateral gene transfer and the nature of bacterial innovation. *Nature* 2000, **405**(6784):299–304.



26. Jain R, Rivera MC, Moore JE, Lake JA: **Horizontal gene transfer in microbial genome evolution.** *Theor Popul Biol* 2002, **61**(4):489–495.
27. Andersson JO, Sarchfield SW, Roger AJ: **Gene transfers from Nanoarchaeota to an ancestor of diplomonads and parabasalids.** *Mol Biol Evol* 2005, **22**(1):85–90.
28. Huang JL, Mullanpudi N, Sicheritz-Ponten T, Kissinger JC: **A first glimpse into the pattern and scale of gene transfer in the Apicomplexa.** *Int J Parasitol* 2004, **34**(3):265–274.
29. Ricard G, McEwan NR, Dutilh BE, Jouany JP, Macheboeuf D, Mitsumori M, McIntosh FM, Michalowski T, Nagamine T, Nelson N, et al: **Horizontal gene transfer from Bacteria to rumen Ciliates indicates adaptation to their anaerobic, carbohydrates-rich environment.** *Bmc Genomics* 2006, **7**:22.
30. Hackett JD, Yoon HS, Soares MB, Bonaldo MF, Casavant TL, Scheetz TE, Nosenko T, Bhattacharya D: **Migration of the plastid genome to the nucleus in a peridinin dinoflagellate.** *Curr Biol* 2004, **14**(3):213–218.
31. Dagan T, Martin W: **Ancestral genome sizes specify the minimum rate of lateral gene transfer during prokaryote evolution.** *Proc Natl Acad Sci U S A* 2007, **104**(3):870–875.
32. Snel B, Bork P, Huynen MA: **Genomes in flux: The evolution of archaeal and proteobacterial gene content.** *Genome Res* 2002, **12**(1):17–25.
33. Huang JL, Mullanpudi N, Lancto CA, Scott M, Abrahamsen MS, Kissinger JC: **Phylogenomic evidence supports past endosymbiosis, intracellular and horizontal gene transfer in *Cryptosporidium parvum*.** *Genome Biol* 2004, **5**(11):R88.
34. Philippe H, Zhou Y, Brinkmann H, Rodrigue N, Delsuc F: **Heterotachy and long-branch attraction in phylogenetics.** *BMC Evol Biol* 2005, **5**:50.
35. Hanten JJ, Pierce SK: **Synthesis of several light-harvesting complex I polypeptides is blocked by cycloheximide in symbiotic chloroplasts in the sea slug, *Elysia chlorotica* (Gould): A case for horizontal gene transfer between alga and animal?** *Biol Bull* 2001, **201**(1):34–44.
36. Wooldridge SA: **A new conceptual model for the warm-water breakdown of the coral-algae endosymbiosis.** *Mar Freshw Res* 2009, **60**(6):483–496.
37. Petherick A: **Salamander's egg surprise.** *Nature* 2010, **466**(7307):675–675.
38. Rumpho ME, Worful JM, Lee J, Kannan K, Tyler MS, Bhattacharya D, Moustafa A, Manhart JR: **Horizontal gene transfer of the algal nuclear gene *psbO* to the photosynthetic sea slug *Elysia chlorotica*.** *Proc Natl Acad Sci U S A* 2008, **105**(46):17867–17871.
39. Habetha M, Bosch TCG: **Symbiotic Hydra express a plant-like peroxidase gene during oogenesis.** *J Exp Biol* 2005, **208**(11):2157–2164.
40. Pierce SK, Fang X, Schwartz JA, Jiang X, Zhao W, Curtis NE, Kocot KM, Yang B, Wang J: **Transcriptomic Evidence for the Expression of Horizontally Transferred Algal Nuclear Genes in the Photosynthetic Sea Slug.** *Molecular biology and evolution: Elysia chlorotica*; 2012.
41. Stiller JW, Huang JL, Ding Q, Tian J, Goodwillie C: **Are algal genes in nonphotosynthetic protists evidence of historical plastid endosymbioses?** *Bmc Genomics* 2009, **10**:484.
42. Bodyl A: **Do plastid-related characters support the chromalveolate hypothesis?** *J Phycol* 2005, **41**(3):712–719.
43. Tyler BM, Tripathy S, Zhang XM, Dehal P, Jiang RHY, Aerts A, Arredondo FD, Baxter L, Bensasson D, Beynon JL, et al: **Phytophthora genome sequences uncover evolutionary origins and mechanisms of pathogenesis.** *Science* 2006, **313**(5791):1261–1266.
44. Hannaert V, Saavedra E, Duffieux F, Szikora JP, Rigden DJ, Michels PAM, Opperdoes FR: **Plant-like traits associated with metabolism of Trypanosoma parasites.** *Proc Natl Acad Sci U S A* 2003, **100**(3):1067–1071.
45. El-Sayed NM, Myler PJ, Blandin G, Berriman M, Crabtree J, Aggarwal G, Caler E, Renauld H, Worthey EA, Hertz-Fowler C, et al: **Comparative genomics of trypanosomatid parasitic protozoa.** *Science* 2005, **309**(5733):404–409.
46. Rogers M, Keeling PJ: **Lateral transfer and compartmentalization of Calvin cycle enzymes of plants and algae.** *J Mol Evol* 2004, **58**(4):367–375.
47. Andersson JO, Roger AJ: **A cyanobacterial gene in nonphotosynthetic protists - An early chloroplast acquisition in eukaryotes?** *Curr Biol* 2002, **12**(2):115–119.
48. Whitaker JW, McConkey GA, Westhead DR: **The transferome of metabolic genes explored: analysis of the horizontal transfer of enzyme encoding genes in unicellular eukaryotes.** *Genome Biol* 2009, **10**(4):R36.
49. Oboznik M, Green BR: **Mosaic origin of the heme biosynthesis pathway in photosynthetic eukaryotes.** *Mol Biol Evol* 2005, **22**(12):2343–2353.
50. Richards TA, Dacks JB, Jenkinson JM, Thornton CR, Talbot NJ: **Evolution of filamentous plant pathogens: Gene exchange across eukaryotic kingdoms.** *Curr Biol* 2006, **16**(18):1857–1864.
51. Richards TA, Soanes DM, Foster PG, Leonard G, Thornton CR, Talbot NJ: **Phylogenomic analysis demonstrates a pattern of rare and ancient horizontal gene transfer between plants and fungi.** *Plant Cell* 2009, **21**(7):1897–1911.
52. Hotopp JCD, Clark ME, Oliveira DCSG, Foster JM, Fischer P, Torres MC, Giebel JD, Kumar N, Ishmael N, Wang SL, et al: **Widespread lateral gene transfer from intracellular bacteria to multicellular eukaryotes.** *Science* 2007, **317**(5845):1753–1756.
53. Klasson L, Kambris Z, Cook PE, Walker T, Sinkins SP: **Horizontal gene transfer between Wolbachia and the mosquito *Aedes aegypti*.** *Bmc Genomics* 2009, **10**:33.
54. Nikoh N, Tanaka K, Shibata F, Kondo N, Hizume M, Shimada M, Fukatsu T: **Wolbachia genome integrated in an insect chromosome: Evolution and fate of laterally transferred endosymbiont genes.** *Genome Res* 2008, **18**(2):272–280.
55. Hotopp JCD: **Horizontal gene transfer between bacteria and animals.** *Trends in Genetics* 2011, **27**(4):157–163.
56. Denker E, Bapteste E, Le Guyader H, Manuel M, Rabet N: **Horizontal gene transfer and the evolution of cnidarian stinging cells.** *Curr Biol* 2008, **18**(18):R858–R859.
57. Chapman JA, Kirkness EF, Simakov O, Hampson SE, Mitros T, Weinmaier T, Rattei T, Balasubramanian PG, Borman J, Busam D, et al: **The dynamic genome of hydra.** *Nature* 2010, **464**(7288):592–596.
58. Gladyshev EA, Meselson M, Arkipova IR: **Massive horizontal gene transfer in bdelloid rotifers.** *Science* 2008, **320**(5880):1210–1213.
59. Moran NA, Jarvik T: **Lateral transfer of genes from fungi underlies carotenoid production in aphids.** *Science* 2010, **328**(5978):624–627.
60. Craig JP, Bekal S, Hudson M, Domier L, Niblack T, Lambert KN: **Analysis of a horizontally transferred pathway involved in vitamin B(6) biosynthesis from the soybean cyst nematode *Heterodera glycines*.** *Mol Biol Evol* 2008, **25**(10):2085–2098.
61. Mayer WE, Schuster LN, Bartelmes G, Dieterich C, Sommer RJ: **Horizontal gene transfer of microbial cellulases into nematode genomes is associated with functional assimilation and gene turnover.** *BMC Evol Biol* 2011, **11**:13.
62. Danchin EGJ, Rosso MN, Vieira P, de Almeida-Engler J, Coutinho PM, Henrissat B, Abad P: **Multiple lateral gene transfers and duplications have promoted plant parasitism ability in nematodes.** *Proc Natl Acad Sci U S A* 2010, **107**(41):17651–17656.
63. Pace JK, Gilbert C, Clark MS, Feschotte C: **Repeated horizontal transfer of a DNA transposon in mammals and other tetrapods.** *Proc Natl Acad Sci U S A* 2008, **105**(44):17023–17028.
64. Armbrust EV, Berges JA, Bowler C, Green BR, Martinez D, Putnam NH, Zhou SG, Allen AE, Apt KE, Bechner M, et al: **The genome of the diatom *Thalassiosira pseudonana*: Ecology, evolution, and metabolism.** *Science* 2004, **306**(5693):79–86.
65. Hooper SD, Berg OG: **Duplication is more common among laterally transferred genes than among indigenous genes.** *Genome Biol* 2003, **4**(8):R48.
66. Sun G, Yang Z, Kosch T, Summers K, Huang J: **Evidence for acquisition of virulence effectors in pathogenic chytrids.** *BMC Evol Biol* 2011, **11**:195.
67. Novakova E, Moran NA: **Diversification of genes for carotenoid biosynthesis in aphids following an ancient transfer from a fungus.** *Mol Biol Evol* 2012, **29**(1):313–323.
68. Belbahri L, Calmin G, Mauch F, Andersson JO: **Evolution of the cutinase gene family: evidence for lateral gene transfer of a candidate *Phytophthora* virulence factor.** *Gene* 2008, **408**(1–2):1–8.
69. Lawrence JG, Ochman H: **Molecular archaeology of the *Escherichia coli* genome.** *Proc Natl Acad Sci U S A* 1998, **95**(16):9413–9417.
70. Koepsell H, Endou H: **The SLC22 drug transporter family.** *Pflugers Archiv-European J Physiology* 2004, **447**(5):666–676.
71. Eraly SA, Monte JC, Nigam SK: **Novel slc22 transporter homologs in fly, worm, and human clarify the phylogeny of organic anion and cation transporters.** *Physiological Genomics* 2004, **18**(1):12–24.
72. Takanaga H, Mackenzie B, Hediger MA: **Sodium-dependent ascorbic acid transporter family SLC23.** *Pflugers Archiv-European J Physiology* 2004, **447**(5):677–682.
73. Murer H, Forster I, Biber J: **The sodium phosphate cotransporter family SLC34.** *Pflugers Archiv-European J Physiology* 2004, **447**(5):763–767.
74. Santoro B, Liu DT, Yao H, Bartsch D, Kandel ER, Siegelbaum SA, Tibbs GR: **Identification of a gene encoding a hyperpolarization-activated pacemaker channel of brain.** *Cell* 1998, **93**(5):717–729.

75. Kallberg Y, Oppermann U, Jornvall H, Persson B: **Short-chain dehydrogenase/reductase (SDR) relationships: A large family with eight clusters common to human, animal, and plant genomes.** *Protein Sci* 2002, **11**(3):636–641.
76. Rokas A: **The molecular origins of multicellular transitions.** *Curr Opin Genet Dev* 2008, **18**(6):472–478.
77. Rokas A: **The Origins of multicellularity and the early history of the genetic toolkit for animal development.** *Annu Rev Genet* 2008, **42**:235–251.
78. O'Brien EA, Koski LB, Zhang Y, Yang LS, Wang E, Gray MW, Burger G, Lang BF: **TBestDB: a taxonomically broad database of expressed sequence tags (ESTs).** *Nucleic Acids Res* 2007, **35**:D445–D451.
79. Huang XQ, Madan A: **CAP3: A DNA sequence assembly program.** *Genome Res* 1999, **9**(9):868–877.
80. Rice P, Longden I, Bleasby A: **EMBOSS: The European molecular biology open software suite.** *Trends in Genetics* 2000, **16**(6):276–277.
81. Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins DG: **The CLUSTAL\_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools.** *Nucleic Acids Res* 1997, **25**(24):4876–4882.
82. Larkin MA, Blackshields G, Brown NP, Chenna R, McGettigan PA, McWilliam H, Valentin F, Wallace IM, Wilm A, Lopez R, *et al*: **Clustal W and clustal X version 2.0.** *Bioinformatics* 2007, **23**(21):2947–2948.
83. Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O: **New Algorithms and Methods to Estimate Maximum-Likelihood Phylogenies: Assessing the Performance of PhyML 3.0.** *Syst Biol* 2010, **59**(3):307–321.
84. Felsenstein J: *PHYLIP (Phylogeny Inference Package) version 3.65.* Seattle (WA): Department of Genome Sciences, University of Washington; 2005.
85. Keane TM, Creevey CJ, Pentony MM, Naughton TJ, McInerney JO: **Assessment of methods for amino acid matrix selection and their use on empirical data shows that ad hoc assumptions for choice of matrix are not justified.** *BMC Evol Biol* 2006, **6**:29.
86. Muller J, Muller K: **TREEGRAPH: automated drawing of complex tree figures using an extensible tree description format.** *Mol Ecol Notes* 2004, **4**(4):786–788.
87. Stover BC, Muller KF: **TreeGraph 2: Combining and visualizing evidence from different phylogenetic analyses.** *BMC Bioinforma* 2010, **11**:7.

doi:10.1186/1471-2148-12-83

**Cite this article as:** Ni *et al.*: Ancient gene transfer from algae to animals: Mechanisms and evolutionary significance. *BMC Evolutionary Biology* 2012 **12**:83.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
[www.biomedcentral.com/submit](http://www.biomedcentral.com/submit)

