

EphA2-receptor deficiency exacerbates myocardial infarction and reduces survival in hyperglycemic mice

DuSablón *et al.*

ORIGINAL INVESTIGATION

Open Access

EphA2-receptor deficiency exacerbates myocardial infarction and reduces survival in hyperglycemic mice

Augustin DuSablón¹, Susan Kent¹, Anita Coburn² and Jitka Virag^{1*}

Abstract

Background: We have previously shown that EphrinA1/EphA expression profile changes in response to myocardial infarction (MI), exogenous EphrinA1-Fc administration following MI positively influences wound healing, and that deletion of the EphA2 Receptor (EphA2-R) exacerbates injury and remodeling. To determine whether or not ephrinA1-Fc would be of therapeutic value in the hyperglycemic infarcted heart, it is critical to evaluate how ephrinA1/EphA signaling changes in the hyperglycemic myocardium in response to MI.

Methods: Streptozotocin (STZ)-induced hyperglycemia in wild type (WT) and EphA2-receptor mutant (EphA2-R-M) mice was initiated by an intraperitoneal injection of STZ (150 mg/kg) 10 days before surgery. MI was induced by permanent ligation of the left anterior descending coronary artery and analyses were performed at 4 days post-MI. ANOVAs with Student-Newman Keuls multiple comparison post-hoc analysis illustrated which groups were significantly different, with significance of at least $p < 0.05$.

Results: Both WT and EphA2-R-M mice responded adversely to STZ, but only hyperglycemic EphA2-R-M mice had lower ejection fraction (EF) and fractional shortening (FS). At 4 days post-MI, we observed greater post-MI mortality in EphA2-R-M mice compared with WT and this was greater still in the EphA2-R-M hyperglycemic mice. Although infarct size was greater in hyperglycemic WT mice vs normoglycemic mice, there was no difference between hyperglycemic EphA2-R-M mice and normoglycemic EphA2-R-M mice. The hypertrophic response that normally occurs in viable myocardium remote to the infarct was noticeably absent in epicardial cardiomyocytes and cardiac dysfunction worsened in hyperglycemic EphA2-R-M hearts post-MI. The characteristic interstitial fibrotic response in the compensating myocardium remote to the infarct also did not occur in hyperglycemic EphA2-R-M mouse hearts to the same extent as that observed in the hyperglycemic WT mouse hearts. Differences in neutrophil and pan-leukocyte infiltration and serum cytokines implicate EphA2-R in modulation of injury and the differences in ephrinA1 and EphA6-R expression in governing this are discussed.

Conclusions: We conclude that EphA2-mutant mice are more prone to hyperglycemia-induced increased injury, decreased survival, and worsened LV remodeling due to impaired wound healing.

Keywords: Diabetes, Ischemia, Myocardial infarction, Hyperglycemia, EphrinA1/EphA

* Correspondence: virag@ecu.edu

¹Department of Physiology, Brody School of Medicine, East Carolina University, 600 Moye Blvd, Greenville, NC 6N-98, USA

Full list of author information is available at the end of the article

Background

Diabetes affects nearly 6% of the US population and about 30% of acute coronary syndrome patients hospitalized have diabetes [1]. In 2004, 68% of diabetes-related deaths were associated with heart disease. Morbidity, mortality, and re-infarction rates are 2-4 times higher following MI in diabetic than in non-diabetic subjects, with one-year mortality in this population as high as 50% [2]. Despite the evidence for improved outcomes from cardiovascular disease in the general population over the past three decades, these benefits have not been paralleled in the diabetic population since the diabetic heart is refractory to known cardioprotective approaches. The heart, being a terminally differentiated organ, lacks endogenous regenerative capacity and thus, developing techniques to maximize myocardial salvage post MI continues to be a health care priority.

The Eph receptors and their cognate ligands, the ephrins are the largest family of receptor tyrosine kinases (RTKs) and their signaling has been implicated in various cellular processes during embryogenesis, tumorigenesis, vasculogenesis and in the regulation of inflammation and apoptosis [3–12]. Previous research has shown that exogenous administration of EphrinA1-Fc following an ischemic event reduces cardiac injury by reducing cardiomyocyte death and inflammation, and promoting tissue regeneration [13–15]. To investigate whether or not ephrinA1-Fc also has potential therapeutic value in the hyperglycemic heart, it is first necessary to compare and contrast ephrinA/EphA expression in WT hyperglycemic and normoglycemic myocardium versus hyperglycemic and normoglycemic EphA2-R-M myocardium following an MI. We hypothesize that EphrinA1/EphA signaling is differentially and unfavorably influenced by hyperglycemia in EphA2-R-M mice and that injury, remodeling, and dysfunction are further exacerbated by hyperglycemia.

Our results indicate that in mice lacking a functional EphA2-R, hyperglycemia worsens remodeling and cardiac function post-MI and this is due to an aggravated inflammatory response, poor infarct resolution, and inadequate healing. This is at least in part due to insufficient production of endogenous protective ephrinA1 in both normoglycemic and hyperglycemic WT and EphA2-R-M mice as well as increased EphA6 expression in EphA2-R-M compared to WT mice. We conclude that exogenous administration of EphrinA1 would be of therapeutic value in the hyperglycemic heart and EphA2 is necessary for appropriate signal transduction. Further studies to assess this are in progress.

Methods

Ethical approval

All procedures were approved by the East Carolina University Institutional Animal Care and Use Committee

and the investigation conforms to the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health.

Animals

WT129SF2/J mice (stock #101045) (WT) and WT129S6-Epha2^{tm1Jnu1}/J (stock #006028; a pan-knockout; EphA2-R-M) mice aged 8-12 weeks were used in these experiments. Male mice were housed in individually ventilated cages and animal care was maintained by the Department of Comparative Medicine at The Brody School of Medicine at East Carolina University. Mice were exposed to 12 h/12 h light/dark cycle conditions and received food and water ad libitum.

Blood glucose

WT and EphA2-R-M mice were given an intraperitoneal injection of 150 mg/kg of streptozotocin (STZ) or 0.1 mol/l of citrate buffer. Mice became diabetic (as evidence by polyuria and polydipsia) at least 1 week prior to surgery. Ten days after the STZ/citrate injection, but prior to surgery, mice were fasted for 4 hours and blood glucose levels were measured (OneTouch Ultra glucometer, Lifescan, CA, USA). Mice were considered hyperglycemic if blood glucose was greater than 200 mg/dl. There were four groups of control mice and 4 groups of infarcted mice (WT citrate, WT STZ, EphA2-R-M citrate, and EphA2-R-M STZ) from which tissue was collected for analysis 4 days post-MI.

Surgical procedure, cardiac function and tissue collection

Myocardial infarction was induced by permanently ligating the left coronary artery in anesthetized, mechanically ventilated mice. Briefly, male WT and EphA2-R-M mice (8-12 weeks) were anesthetized with an intraperitoneal (IP) injection of 20 µl/g body weight Avertin (20 mg/ml), a left thoracotomy performed, and the left anterior descending coronary artery was permanently occluded using an 8-0 suture. The rib cage, muscle, and skin were then closed with a 6-0 suture. Uninjured sham controls underwent the entire procedure including suture pass under the coronary [13,16].

Echocardiography was performed on conscious mice at 4 days post-MI. A VisualSonics Vevo 2100 diagnostic ultrasound, using M-mode and 30 MHz probe, was used to obtain LV dimensions in diastole and systole. End-diastolic measurements of interventricular septal thickness (IVSd), left ventricular posterior wall thickness (LVPWd), and left ventricular internal diameter (LVIDd) were obtained at the point of maximal LV diastolic dimension. End-systolic dimensions (IVSs, LVPWs, and LVIDs) were measured at the time of most anterior systolic excursion of the LVPW associated with minimal chamber dimension. Average measurements were calculated using the

leading-edge technique of 3- to 5-consecutive sinus beats. Ejection fraction (EF) was calculated from LV dimensions above using the following formula: $(LVIDd^3 - LVIDs^3)/LVIDd^3 \times 100\%$. FS was calculated using the formula $(LVIDs-LVIDd)/LVIDd \times 100\%$ [17].

At the time of sacrifice, mice were anesthetized with an IP injection of 0.1 mL pentobarbital (390 mg/mL) and a pneumothorax was performed. Hearts to be used for histology and immunohistochemistry were arrested in diastole using cold KCl (30 mM), excised, rinsed in PBS, and immersed in zinc fixative. For RNA and protein analyses, the right ventricle was dissected away from the heart and discarded while the remaining left ventricle was snap frozen in liquid nitrogen.

Histology, morphometry, and immunohistochemistry

Tissue was harvested and immersed in zinc fixative, then processed and embedded in paraffin as per routine procedures. For immunohistochemical staining and morphometric measurements, whole hearts were transversely sectioned into 4 slices of equal thickness, processed in an automated tissue processor (TP1020, Leica, Nußloch, Germany), and embedded in paraffin (MICROM EC350, Richard-Allan Scientific, Kalamazoo, MI, USA) [18,19]. Blocks were then sectioned into 5µm sections and mounted on Superfrost Plus glass slides for histologic or immunohistochemical staining.

Morphometric measurements of the left ventricle, infarct area, and chamber area were performed on tissue sections 4 days post-MI (n = 7-12/group). Images of 4 hematoxylin and eosin (H&E)-stained sections of each heart were taken at $\times 20$ magnification using a DP70 digital camera. Myocyte cross-sectional area (MCSA) was measured in 3-8 cardiomyocytes with centrally located nuclei in each of 6 images (600 \times) in the endocardium and epicardium (n = 3-5/group) using Scion imaging software (Scion Corporation, Frederick, MD, USA).

Slides stained with picosirius red for fibrillar collagen and fast green for contrast were used to quantify interstitial fibrosis [20]. Adobe Photoshop software was used to count the red collagen fibril pixels which were expressed as a percentage of the total number of pixels (total = green + red - white) in five images at 400 \times taken in each of 2 cross sections per heart remote to the infarct but in the same plane.

Tissue sections were deparaffinized in xylene and endogenous peroxidases quenched with 3% H₂O₂ in methanol. As described elsewhere previously, slides were rinsed in PBS and incubated with Ly6G (BD Biosciences, #550291) to assess neutrophil infiltration, CD45 (BD Biosciences; #550539) for macrophage density [13,19-22]. Isolectin B4 (Vector Labs; B-1205) or CD31 (BD Biosciences, #553371) was used to measure capillary density in uninjured control and infarcted hearts respectively

[19]. Slides were incubated with appropriate biotinylated secondary antibodies and then with Avidin Biotin Complex (Vector Labs PK-6100). The reaction product was visualized with DAB (Vector, SK-4100), counterstained with methyl green, dehydrated in xylene, and slides were coverslipped.

ELISA

A mouse cytokine proteome profiler array (R&D #ARY006) was used to compare levels of 40 serum cytokines as per manufacturer's protocol. Forty microliters from each of 5 different samples were combined for each group. The samples were run in duplicate and data are expressed as relative expression (pixel density = relative units) with levels appropriately normalized to internal reference spot controls.

qRT-PCR

Whole left ventricles from mice 4 days post-MI were homogenized using Trizol for RNA isolation. Purification was performed using the Qiagen RNeasy kit. cDNA was made for each sample using a high capacity cDNA kit. Real-time PCR (qRT-PCR) performed using an Applied Biosystems thermocycler. TaqMan primers were obtained from Applied Biosciences (ephrinA1: Mm00438660_m1, EphA1: Mm00445804_m1, EphA3: Mm00580743_m1, EphA4: Mm00433056_m1, EphA5: Mm00433074_m1, EphA6: Mm00433094_m1, EphA7: Mm00833876_m1, GAPDH: Mm99999915_g1). All samples were run in triplicate and a reaction mixture of 10 µl (100 ng RNA) was amplified using recommended conditions from Applied Biosciences. Gene expression was normalized to expression of GAPDH. Fluorescence data were analyzed using the $\Delta\Delta$ Ct method.

Western blotting

Whole left ventricles of uninjured sham control hearts and hearts 4 days post-MI from each of the 8 groups of mice were homogenized in a lysis buffer containing 50 mM Hepes, 10 mM EDTA, 100 mM NaF, 50 mM sodium pyrophosphate, 1% protease, and 1% phosphatase inhibitors. The Bradford Assay was used to quantify the amount of protein. Western blotting was performed on a 4-12% gradient Bis-Tris gel (BioRad) in 1 \times Mops running buffer.

Fifty micrograms of sample (n = 3/group) was loaded per well. The gel was run for 1 h at 155 V, and transferred onto pure nitrocellulose membranes (BioRad). Antibodies: GAPDH (Millipore, #MAB374), NF- κ B (Santa Cruz, #sc-372), EphA6-R (SantaCruz #25740), MMP-9 (R&D, BAF909), p-mTOR (Cell Signaling, #5536), mTOR (Cell Signaling, #2983), and ephrinA1 (Santa Cruz, #sc-911) were followed by appropriate secondary antibodies. All blots were detected with Immun-Star Horseradish

peroxidase chemiluminescence (Bio-Rad, Hercules, CA) and imaged on an Alpha Innotech imaging system (ProteinSimple, San Jose, CA). Densitometry was performed using Image J 1.42 software (NIH, Bethesda, MD) and the intensity of each protein was normalized to GAPDH.

Statistics

ANOVA (analysis of variance) with Student-Newman Keuls multiple comparison post-hoc analysis illustrated which groups were statistically different, with significance of at least $p < 0.05$.

Results

Survival

The Kaplan-Meier graph in Figure 1 depicts the survival rates during the first 4 days post-MI. Mice subjected to STZ + MI had lower cumulative survivals than animals with only MI. WT ($n = 5$) and EphA2-R-M ($n = 6$) control animals had a cumulative survival of 100% and 83.3% 4 days post-MI, respectively. WT STZ ($n = 17$) had a cumulative survival of 70.6% 4 days post-MI and EphA2-R-M STZ ($n = 26$) had a cumulative survival of 50% after MI.

Infarct size

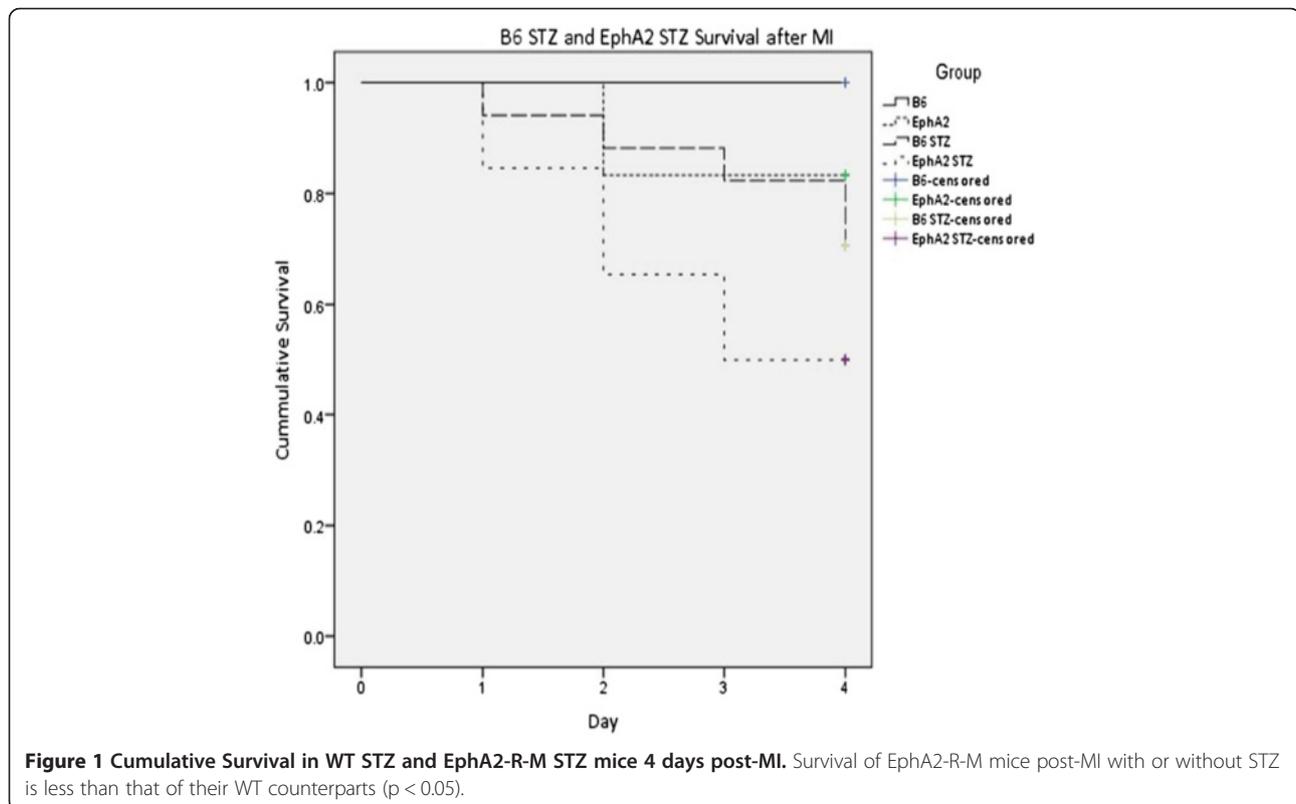
Infarct size 4 days post-MI is shown in Figure 2. 4 days post-MI, the infarct size in WT mouse hearts ($n = 12$) was $37.5 \pm 4.3\%$ of the left ventricle compared with $49.5 \pm$

3.3% in EphA2-R-M mice ($n = 10$). In hyperglycemic WT mice, infarct size at 4 days post-MI was $68\% \pm 7.7\%$ ($n = 8$) compared with $51.5 \pm 6.2\%$ in EphA2-R-M ($n = 9$) mice. There were no differences between LV and chamber area at 4 days post-MI compared to hyperglycemic mice of both WT and EphA2-R-M mice at 4 days post-MI. There were no differences in endocardial myocyte cross-sectional area (MCSA) among the 8 groups studied. STZ-treated groups showed a nonsignificant trend to have decreased epicardial MCSA. Epicardial cardiomyocytes in EphA2-R-M mice demonstrated a non-significant trend to hypertrophy post-MI (263.9 ± 15.4 vs 315.0 ± 30.4) but this was blunted in the STZ-treated EphA2-R-M mice (203.8 ± 8.9 vs 222.3 ± 21.1). Similarly, there was an increase in the interstitial fibrosis in the remote myocardium in normoglycemic mice post-MI and this increased nearly 2-fold in hyperglycemic WT mice but did not change in hyperglycemic EphA2-R-M mice (representative images Figure 3A-D; numerical Figure 3E).

Ly6-G⁺ neutrophil density

STZ

Ly6-G⁺ cell counts to determine neutrophil density in control and STZ-treated WT and EphA2-R-M mouse hearts is shown in Figure 4. Neutrophil density in normoglycemic WT ($n = 6$) hearts was 0.21 ± 0.04 cells/ 0.1 mm^2 compared with the normoglycemic EphA2-R-M ($n = 3$) hearts with 3.48 ± 0.29 cells/ 0.1 mm^2 ($p < 0.01$). In



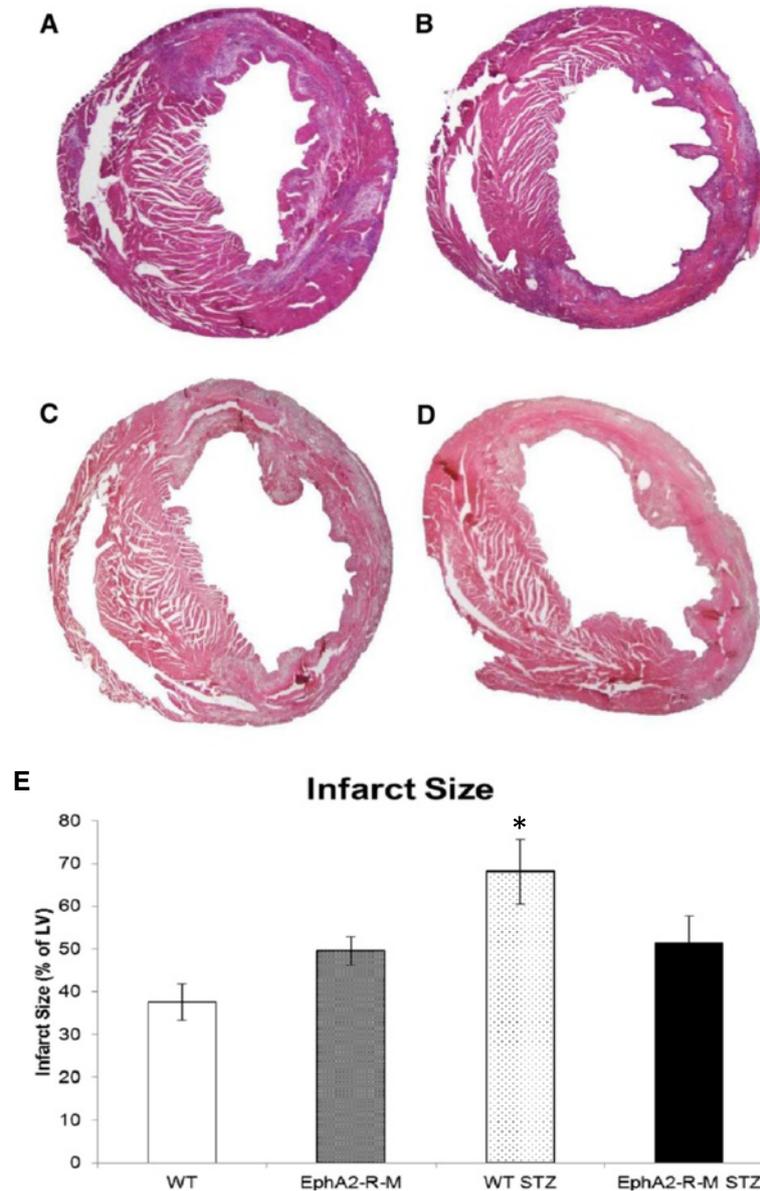


Figure 2 Infarct size in WT and EphA2-R-M Normo- and Hyperglycemic mice. Representative images of H&E stains (x20) of normoglycemic WT mouse heart (A) and EphA2-R-M (B), and hyperglycemic WT (C) and EphA2-R-M (D) mouse hearts. Infarct size (E) in WT mice was 45% larger in WT hyperglycemic mice compared with normoglycemic mice ($p < 0.01$). EphA2-R-M mice had slightly with slightly larger infarcts but this was not exacerbated by hyperglycemia (E).

hyperglycemic mouse hearts we observed a neutrophil density of 4.10 ± 0.91 cells/ 0.1 mm^2 in WT STZ ($n = 6$) hearts and 1.24 ± 0.25 cells/ 0.1 mm^2 in EphA2-R-M STZ ($n = 7$) hearts ($p < 0.05$). Neutrophil density in WT STZ was nearly 19-fold higher than WT controls ($p < 0.01$). EphA2-R-M STZ hearts however, had 64% less neutrophils than EphA2-R-M controls ($p < 0.01$).

STZ + MI

Ly6-G⁺ staining for neutrophil density in the infarct zone 4 days post-MI (Figure 4) showed an increase in

EphA2-R-M (70.04 ± 7.86 cells/ 0.1 mm^2 ; $n = 9$) compared with WT (49.38 ± 5.26 cells/ 0.1 mm^2 ; $n = 10$) mouse hearts ($p < 0.05$). Neutrophil density 4 days post-MI was not different between the WT + STZ mouse hearts (92.89 ± 23.34 cells/ 0.1 mm^2) and the EphA2-R-M + STZ post-MI (91.89 ± 10.25 cells/ 0.1 mm^2). In WT mice, MI alone increased neutrophil infiltration 103% over infarcted EphA2-R-M mice ($p = 0.06$). STZ + MI only increased neutrophils by a non-significant 31% in EphA2-R-M compared to STZ + MI WT hearts ($p = 0.12$).

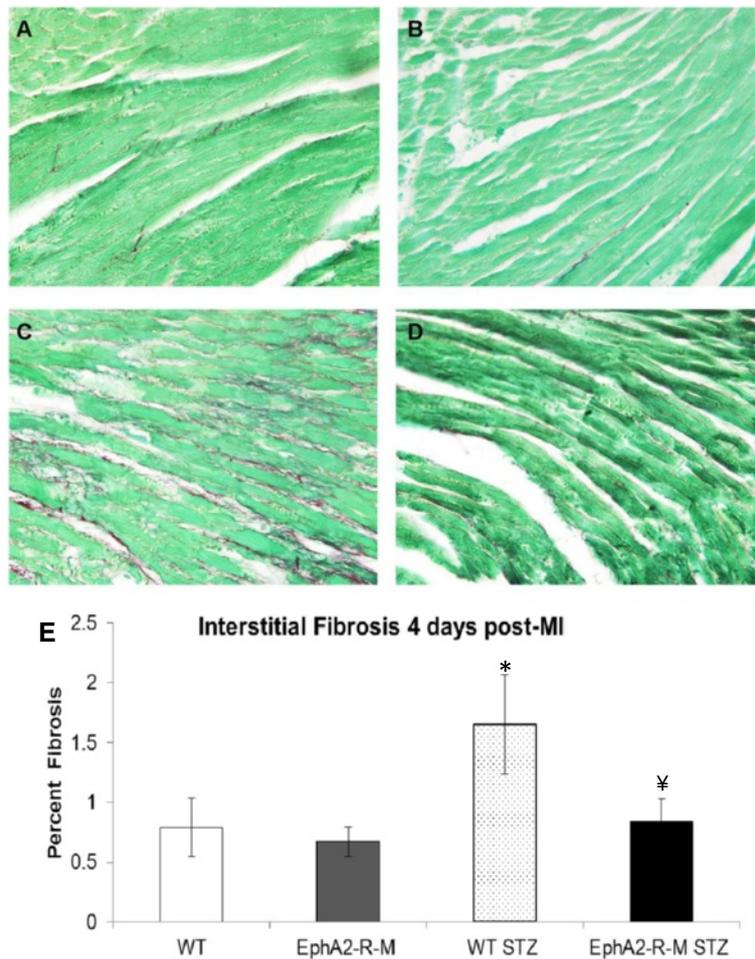


Figure 3 Interstitial Fibrosis. Representative images (x400) of normoglycemic WT mouse heart (A) and EphA2-R-M (B), and hyperglycemic WT (C) and EphA2-R-M (D) mouse hearts stained with picosirius red and fast green. Percent interstitial fibrosis (E) was elevated in hyperglycemic WT mice 4 days post-MI (*, $p < 0.05$) compared to normoglycemic WT hearts and hyperglycemic EphA2-R-M mouse hearts (¥, $p < 0.01$).

CD45⁺ macrophage density STZ

Macrophage density in normoglycemic hearts was no different between WT ($n = 3$) and EphA2-R-M ($n = 10$) (Figure 5). CD45⁺ staining of macrophage infiltration in WT STZ ($n = 6$) hearts was 18.86 ± 1.22 cells/ 0.1 mm^2 compared with the EphA2-R-M STZ ($n = 6$) with 15.85 ± 0.91 cells/ 0.1 mm^2 ($p = 0.07$). There was a 147% increase in macrophage infiltration in WT STZ ($n = 6$) compared with WT controls ($n = 3$) ($p < 0.01$). In EphA2-R-M STZ ($n = 6$), there was an insignificant increase of only 26% in macrophage infiltration compared with EphA2-R-M controls ($p = 0.22$).

STZ + MI

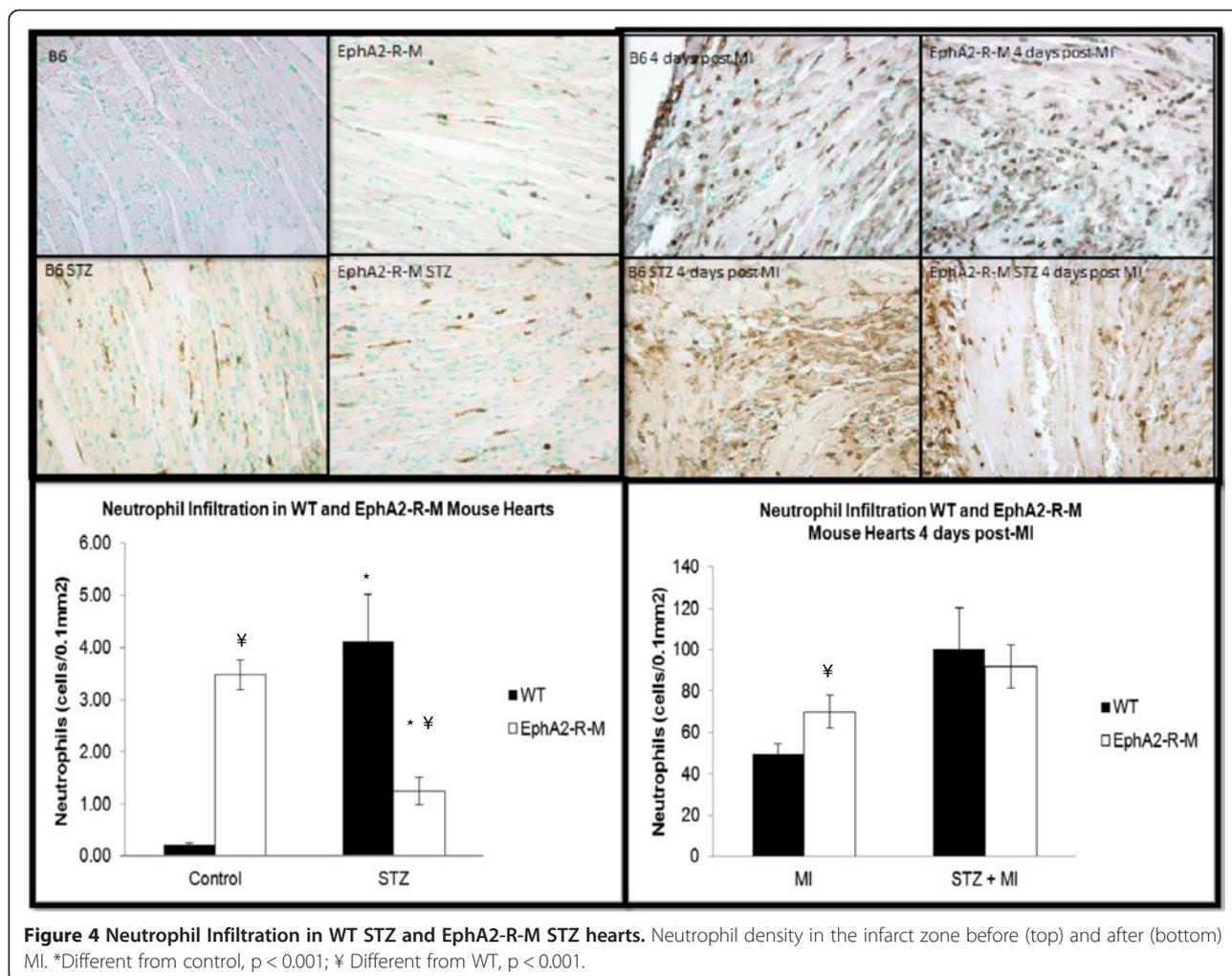
Macrophage density 4 days post-MI (Figure 5) in normoglycemic WT ($n = 7$) hearts was 55.97 ± 5.94 cells/ 0.1 mm^2 compared with the normoglycemic EphA2-R-M

($n = 8$) hearts with 102.76 ± 5.94 cells/ 0.1 mm^2 ($p < 0.001$). Macrophage density was no different after MI in WT STZ ($n = 5$) and EphA2-R-M STZ ($n = 5$) hearts ($p = 0.65$). Macrophage infiltration was 17% less in WT STZ ($n = 5$) hearts after MI compared with WT ($n = 7$) hearts ($p = 0.34$). However, EphA2-R-M STZ hearts post-MI ($n = 5$) had 64% less macrophages than EphA2-R-M hearts after MI ($n = 8$) ($p < 0.01$).

Serum cytokines

Serum cytokine levels are shown in Figure 6. Of the 40 cytokines tested by the array, only 10 were expressed in the samples tested and 4 of these, TREM, TNF- α , IL-1 α , and MIG were only detected in the EphA2-R-M 4d + STZ group (data not shown).

C5a and sICAM-1 were present in all groups (panels A - D) whereas M-CSF, MCP-1, KC, and TIMP-1 were only detected in injured hearts (panels E- H).



Capillary density STZ

Endocardial capillary density in WT control ($n = 6$) hearts was 167 ± 18 vessels per 400x high power field and 121 ± 8 vessels per 400x high power field in EphA2-R-M control ($n = 5$) hearts ($p < 0.05$). In WT STZ hearts ($n = 5$), endocardial capillary density was reduced by 82% compared with control hearts ($p < 0.001$). EphA2-R-M STZ hearts ($n = 5$) observed a decrease in capillary density in the endocardium by 89% compared with WT control hearts ($p < 0.001$). EphA2-R-M STZ ($n = 5$) hearts had 54% less capillaries in the endocardium than WT STZ ($n = 5$) hearts ($p < 0.01$). These results are shown in Figure 7A.

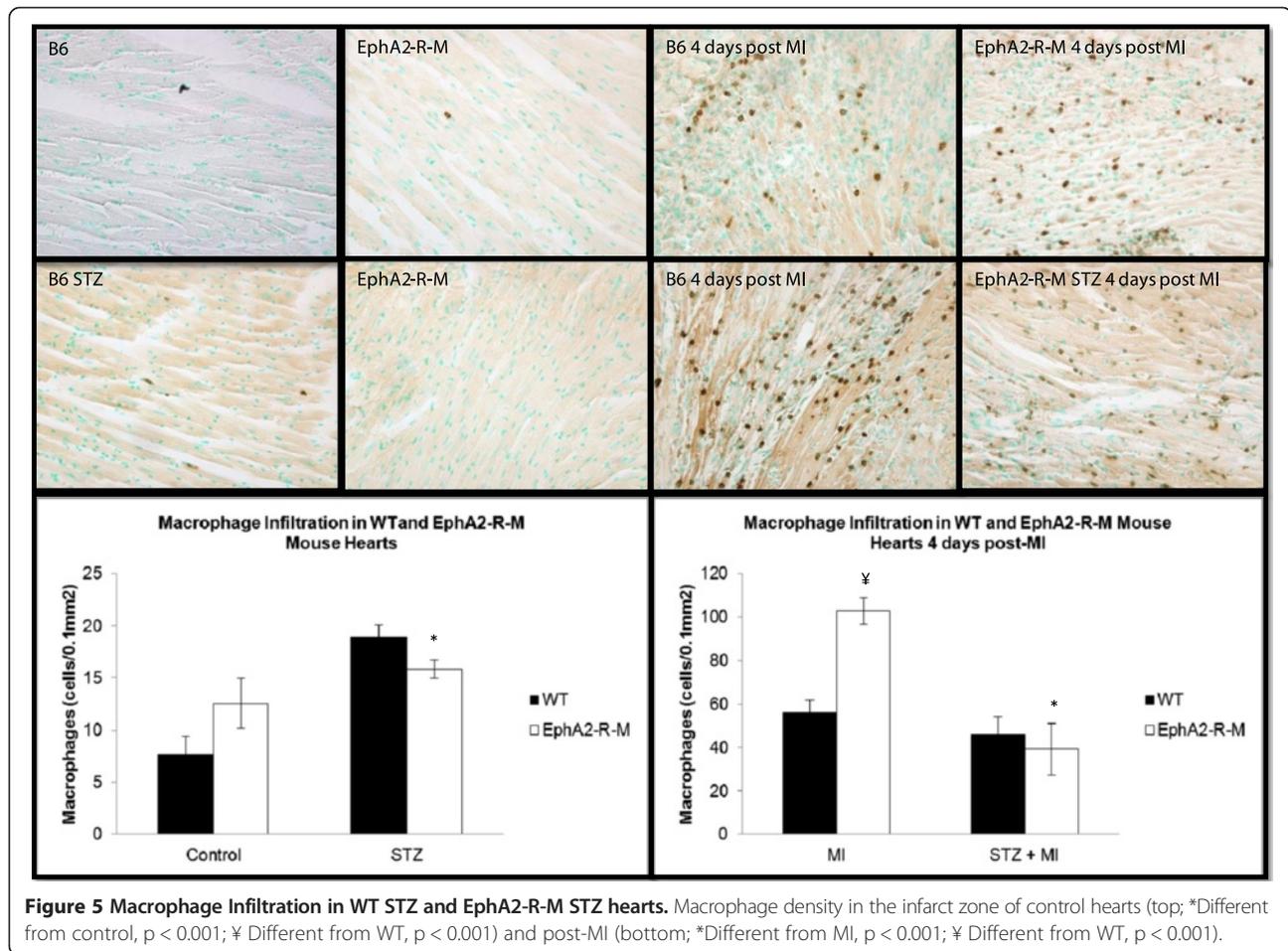
STZ + MI

Endocardial CD31⁺ staining 4 days post-MI showed that capillary density was reduced by 63% in normoglycemic EphA2-R-M ($n = 6$) hearts compared with WT ($n = 4$) ($p < 0.05$). After MI, WT STZ ($n = 5$) had an increase in capillary density of 109% compared with WT control

($n = 4$) hearts. ($p < 0.05$). EphA2-R-M STZ ($n = 4$) had 53% less capillaries than EphA2-R-M control ($n = 6$) hearts after MI ($p < 0.05$). EphA2-R-M STZ ($n = 4$) had 92% less capillary density than WT STZ ($n = 5$) after infarction ($p < 0.01$). These results are depicted in Figure 7A.

Epicardial capillary density in WT control ($n = 6$) hearts was 108.97 ± 12.98 vessels per 400x high power field and 100.90 ± 11.71 vessels per 400x high power field in EphA2 control ($n = 5$) hearts ($p = 0.65$). In WT STZ hearts ($n = 5$), epicardial capillary density was reduced by 80.3% compared with WT control hearts ($p < 0.001$). EphA2 STZ hearts ($n = 5$) observed a decrease in capillary density in the epicardium by 86% compared with EphA2-R-M control hearts ($p < 0.001$). EphA2 STZ ($n = 5$) hearts had 33.3% less capillaries in the epicardium than WT STZ ($n = 5$) hearts ($p = 0.15$). These results are shown in Figure 7B.

Epicardium CD31⁺ staining 4 days post-MI was 27.38 ± 1.99 vessels per 400x high power field in WT control ($n = 4$) hearts and 9.04 ± 0.73 vessels per 400x high



power field in EphA2 ($n = 6$) control hearts after MI ($p < 0.01$). WT STZ ($n = 5$) had an increase in epicardial capillary density of 46% compared with WT ($n = 4$) hearts after infarction ($p < 0.01$). EphA2 STZ ($n = 4$) had a 26.4% decrease in epicardial capillary density compared with EphA2 ($n = 6$) control hearts after MI ($p = 0.25$). After infarction, EphA2 STZ ($n = 4$) hearts had 83.4% less epicardial capillaries than WT STZ ($n = 5$) hearts ($p < 0.001$). These results are depicted in Figure 7B.

Echocardiography

STZ

Echocardiographic parameters for each of the 8 groups are presented in Table 1. Although there was no difference in EF between WT and EphA2-R-M mice, FS was depressed in EphA2-R-M mice and this difference was also present in hyperglycemic mice. Hyperglycemic EphA2-R-M mice also had reduced HR while WT mice had increased SBP and DBP. There were no differences in LV mass between uninjured normoglycemic and hyperglycemic EphA2-R-M and WT mice.

STZ + MI

EF and FS in both WT and EphA2-R-M mice 4 days post-MI were not different from their respective uninjured controls but diastolic diameter and volume as well as systolic volume are elevated in WT mice 4 days post-MI compared with uninjured control. Hyperglycemic WT and EphA2-R-M mice had depressed cardiac function 4 days post-MI compared with respective normoglycemic infarcted mice. Systolic diameter and volume were only elevated in WT mice. There was no difference in LV mass in infarcted STZ mice but the EphA2-R-M mice did not exhibit proportionate compensatory hypertrophy demonstrated by the WT mice 4 days post-MI.

qRT-PCR

STZ

EphrinA1 and EphA-R receptor gene expression was quantified using qRT-PCR mRNA levels for WT STZ mice ($n = 4$) and EphA2-R-M STZ mice ($n = 5$) (Figure 8). EphrinA1 ligand expression exhibited a non-significant trend to increase (1.24-fold; $p = 0.08$) in EphA2-R-M

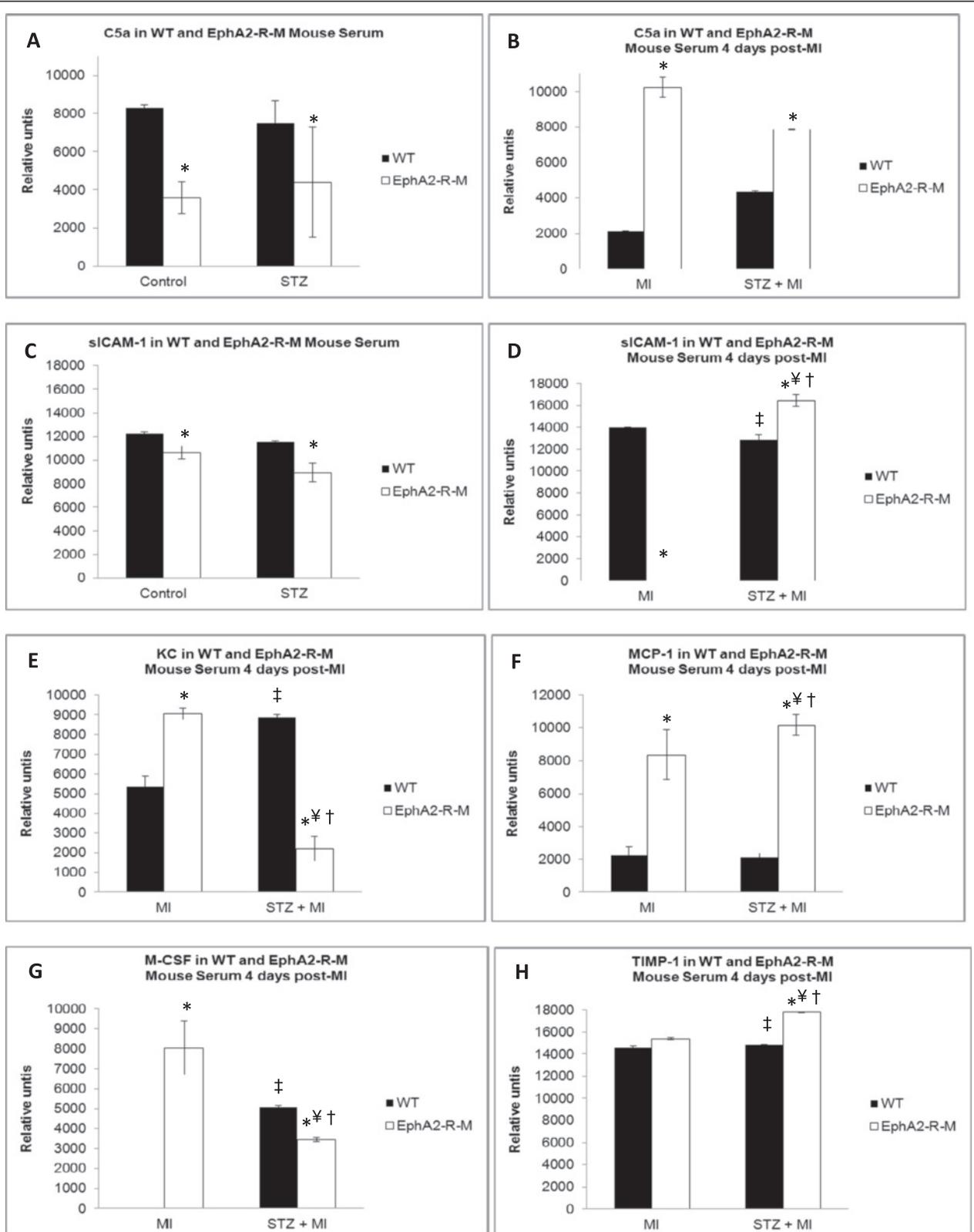


Figure 6 (See legend on next page.)

(See figure on previous page.)

Figure 6 Serum Cytokines in Normoglycemic and Hyperglycemic WT and EphA2-R-M mice. Panel A and C show the differences in C5a and sICAM-1 between normoglycemic and hyperglycemic WT and EphA2-R-M mice. Panel B and D shows how C5a and sICAM-1 change post-MI. Panels E-H show post-MI changes in WT versus EphA2-R-M mice in serum levels of KC, MCP-1, M-CSF, and TIMP-1. ($p < 0.05$: *different from WT, •different from WT STZ, †different from WT 4d, ‡different from EphA2-R-M 4d, †different from WT 4d STZ).

STZ hearts compared with WT STZ hearts. EphA6 also exhibited a non-significant trend to increase (2.01-fold, $p = 0.06$) in EphA2-R-M STZ hearts compared with WT STZ hearts.

STZ + MI

EphrinA1 and Eph Receptor gene expression was also quantified for WT STZ (5) and EphA2-R-M STZ (4) after infarction (Figure 8). Similar to uninjured STZ hearts, ephrinA1 and EphA6 were higher among EphA2-R-M STZ hearts after infarction. EphrinA1 was increased 1.19-fold ($p < 0.05$) and EphA6 was increased nearly 4.5-fold ($p < 0.05$) in EphA2-R-M STZ hearts after infarction.

Western blotting

STZ

Representative blots for ephrinA1 and EphA6-R in WT mice (left) and EphA2-R-M mice (right) are shown in Figure 9. There were no statistically significant differences in expression of NF- κ B, MMP-9, or p-mTOR/mTOR in response to STZ in either group (data not shown).

STZ + MI

Although not significant, there was a trend for reduced ephrinA1 (13%) expression in normoglycemic WT hearts post-MI and increased EphA6-R expression in both normoglycemic and hyperglycemic WT and EphA2-

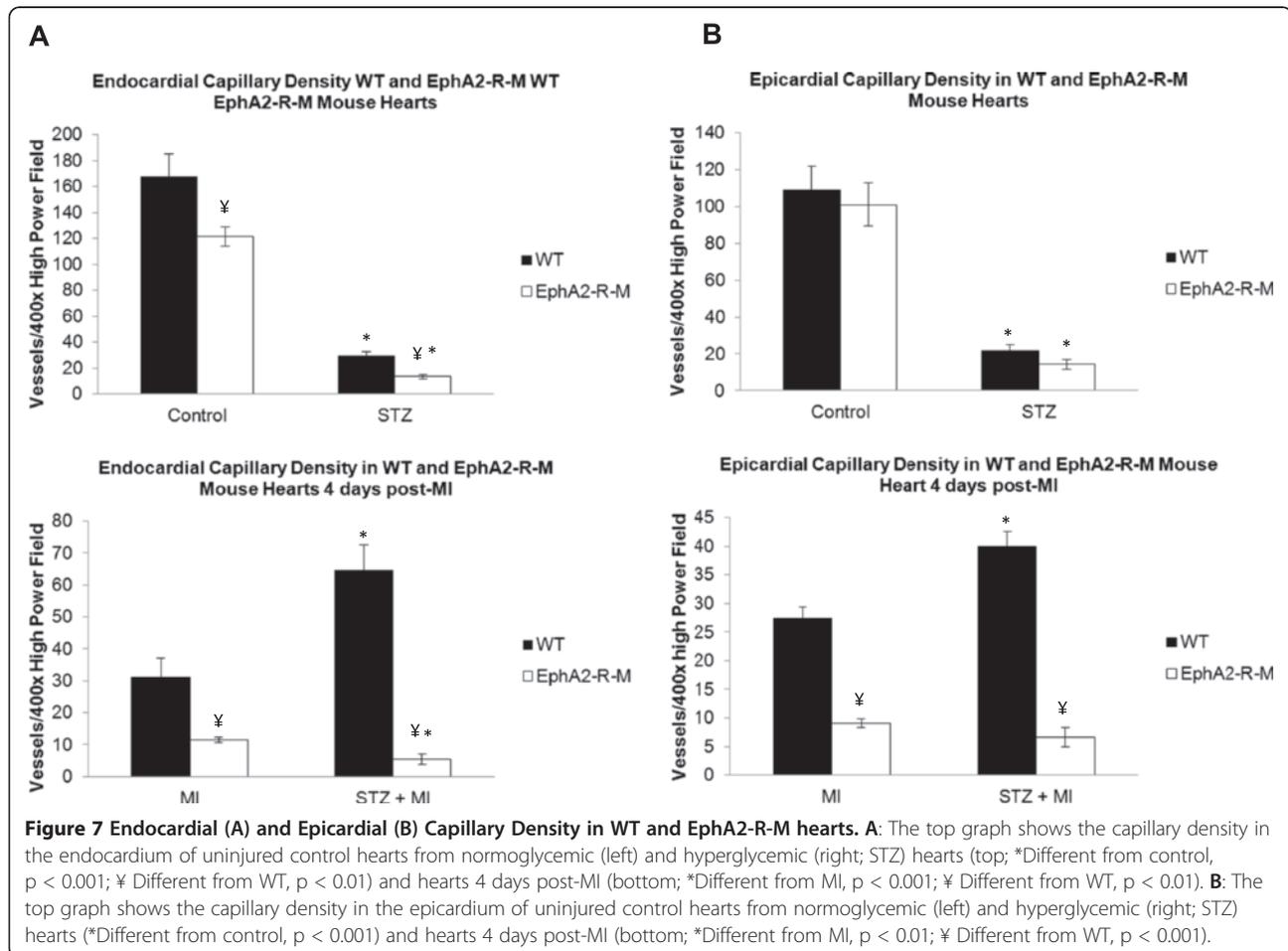


Table 1 Cardiac function and blood pressure parameters

Parameter	EF	FS	LVID _d	LVID _s	LV vol _d	LV vol _s	HR	SBP	DBP	LV mass
WT (n=6)	91 ± 3	60 ± 4	2.7 ± 0.3	1.1 ± 0.1	27.8 ± 7.2	2.5 ± 0.5	619 ± 98	132 ± 4	81 ± 3	102 ± 22
EphA2-R-M (n=9)	82 ± 4	49 ± 4*	2.7 ± 0.2	1.4 ± 0.2	27 ± 5.7	4.8 ± 1.5	636 ± 71	129 ± 4	77 ± 2	71 ± 15
WT STZ (n=8)	88 ± 2	57 ± 3	2.8 ± 0.2	1.2 ± 0.1	30.1 ± 4.7	3.6 ± 0.9	578 ± 98	185 ± 6*	107 ± 11*	69 ± 11
EphA2-R-M STZ (n=18)	78 ± 6	46 ± 6*	2.7 ± 0.2	1.5 ± 0.2	27.8 ± 5.6	6.1 ± 2.4	492 ± 142 [†]	148 ± 4	91 ± 3	60 ± 15
WT 4d (n=7)	77 ± 1	45 ± 5	3.4 ± 0.4*	1.9 ± 0.3	51.0 ± 14.1*	13.5 ± 5.4*	658 ± 91	129 ± 8	73 ± 5	169 ± 51
EphA2-R-M 4d (n=8)	79 ± 3	46 ± 3	3.2 ± 0.1	1.7 ± 0.1	39.5 ± 2.2	8.5 ± 1.5	632 ± 93	126 ± 4	76 ± 3	94 ± 24 [‡]
WT STZ + 4d (n=7)	55 ± 13 [‡]	28 ± 8 [‡]	3.3 ± 0.3	2.4 ± 0.5 [‡]	44.9 ± 10.0	21.0 ± 9.1 [‡]	602 ± 98	127 ± 6	73 ± 7	107 ± 32 [‡]
EphA2-R-M STZ + 4d (n=11)	62 ± 12 [‡]	33 ± 8 [‡]	3.1 ± 0.7	2.1 ± 0.5	39.7 ± 18.6	15.4 ± 9.2	536 ± 80	130 ± 5	69 ± 2	81 ± 40

Echocardiography and blood pressure measures from conscious WT, EphA2-R-M, WT STZ, EphA2-R-M STZ, WT 4d, EphA2 3d, WT STZ + 4d, and EphA2-R-M STZ + 4d mice (p<0.05: *different from WT, [†]different from EphA2-R-M, [•]different from WT STZ, [‡]different from WT 4d, [‡]different from EphA2-R-M 4d).

R-M hearts (11%) post-MI. There were no significant differences in NF-κB, MMP-9, or p-mTOR/mTOR expression between the two groups (data not shown).

Discussion

Cardiomyocyte death post-MI starts immediately after coronary occlusion and occurs downstream from the

occlusion. The therapeutic goal to decrease infarct size to less than 20% of the left ventricle has shown to improve long-term prognosis [23]. Early diagnosis and treatment are critical but presently inadequate. Administering a recombinant protein alone or in conjunction with grafting cells has reduced early injury as well as adverse remodeling and subsequent cardiac dysfunction

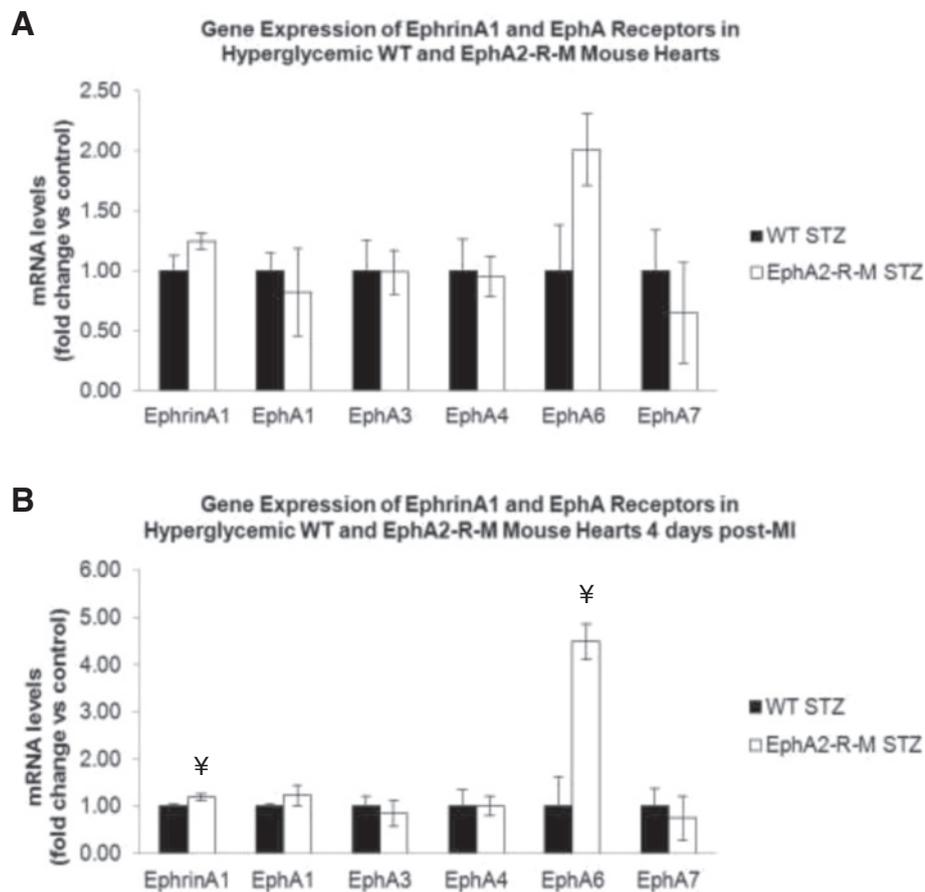


Figure 8 Gene Expression of EphrinA1 and Eph Receptors in WT and EphA2-R-M STZ hearts before (A) and after (B) MI. There were no significant differences between any of the genes tested in uninjured control hearts but ephrinA1 and EphA6-R were elevated in EphA2-R-M hearts post-MI. ¥ Different from WT (p < 0.05).

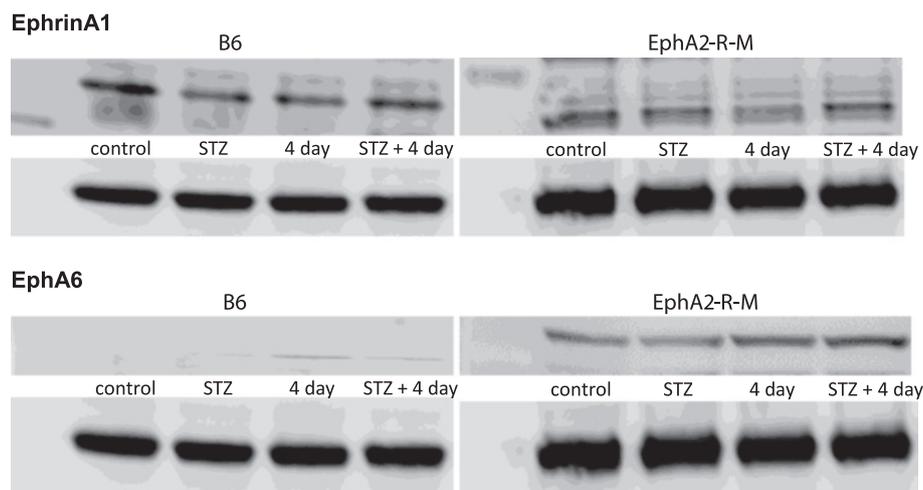


Figure 9 Western blots. Representative samples of left ventricles of uninjured and infarcted hearts from normoglycemic and hyperglycemic WT (left) and EphA2-R-M (right) mice probed with ephrinA1 (top) and EphA6-R (bottom) and normalized to GAPDH.

[24–29]. Clinically, only reperfusion therapy with angioplasty or thrombolysis has been proven to reduce early injury. Although late remodeling cannot be significantly altered, it is managed with pharmacologic agents which sustain ventricular function but this merely delays the progression to heart failure. Current therapeutics, such as early myocardial revascularization after myocardial infarction, are geared toward minimizing cardiac myocyte necrosis and may modulate apoptosis and autophagy [30]. Research that explores the number and type of cells for grafting alone or in combination with gene and protein-based therapeutic strategies are aimed at augmenting the cardiac regenerative potential and promoting tissue salvage. Due to poor reproducibility and efficacy, as well as untoward side effects, none of these experimental paradigms have been successfully translated into clinical therapies. Due to these factors and the narrow window of opportunity between the initial cardiac insult and the cellular demise, developing newer therapies for “cellular resuscitation” are of paramount importance [31–33].

Diabetes has long been associated with greater incidence of myocardial infarction, inadequate healing, and susceptibility to further events. Hyperglycemia, the principal manifestation of all types of diabetes, results in perturbations in cellular metabolism, oxidative stress, exacerbated inflammation, and AGE-modified proteins that culminate in tissue damage, cardiac dysfunction, and impaired wound healing responses [34–42]. Sweeney et al. (2012) observed decreased glucose uptake and increased autophagy after acute I/R injury in STZ hearts, however, long term studies reveal detrimental effects on remodeling that ultimately compromise performance [43].

The EphA2-R has historically been known to regulate apoptosis in developing vasculature and nervous system

and cell death during tumorigenesis [5,44–50], the EphA2-R also modulates vascular permeability, inflammation, and ischemic injury [11,12,14,51–59]. Knockdown of EphA2-R in HUVECs suppresses thrombin-induced phosphorylation of NF- κ B, resulting in decreased ICAM-1 expression [59]. We observed increased sICAM-1 in serum of WT mice post-MI but this is blunted in hyperglycemic mice. Interestingly, sICAM-1 is decreased in hyperglycemic EphA2-R-M mice compared to normoglycemic mice and is not detected post-MI but is increased in hyperglycemic EphA2-R-M post-MI. EphrinA1 and EphA2 are both upregulated in response to inflammation [60–62]. Although we did not detect differences in NF- κ B expression, we did observe decreased ephrinA1 expression in EphA2-R-M mice post-MI, and propose that the deficiency of EphA2-R prevents timely induction of inflammatory cascades necessary for mediating repair. Further, cardiac ephrinA1 expression exhibited a non-significant trend to increase slightly in hyperglycemic WT mice post-MI but not to the extent that they are expressed in uninjured normoglycemic mice, and is thus likely ineffective in eliciting the protective effects known to occur from exogenous administration of ephrinA1-Fc [13–15]. TNF- α , IL-1 α , MIG, and TREM were only detectable in serum of hyperglycemic EphA2-R-M mice, and significant elevation of MCP-1, TIMP-1, and C5a, which typically correlate with the severity of injury, suggests exacerbated wound healing in EphA2-R-M mice [63–67]. Additionally, KC and M-CSF are reduced in hyperglycemic EphA2-R-M mice post-MI, implying poor monocyte adhesion and differentiation [63,68–70]. This is supported by the reduced leukocyte infiltration observed in this group, and although cardiac function is not worsened, the increased mortality in this group indicates that inadequate healing mechanisms acutely result in lethality. Indeed, plasma sICAM-1

level is associated with vascular inflammation and has been shown to correlate with increased risk of cardiovascular events [71,72], providing further support for the continuity of our findings.

We have previously shown that intramyocardial administration of ephrinA1-Fc reduces early injury in non-reperfused myocardium via reduced inflammatory cell infiltration and changes in gene expression of several EphA-Rs. The mechanisms by which this protection occurs can be multifaceted depending on the EphA receptors activated/inhibited and the expression profile of the various cells involved. Goichberg et al. (2012) showed ephrinA1-Fc enhanced motility of EphA2-expressing hCSCs, promoting infarct repair [14]. In adult mouse immortalized HL-1 cardiomyocytes, increased EphA2 expression and reduced phosphorylation following lithocholic acid treatment prevented doxazosin-induced apoptosis via inhibition of SHP-2 [73]. Clearly, the EphA2 receptor plays a vital role in myocardial tissue viability and more studies to understand the signaling pathways involved are warranted.

Expression of the EphA6 receptor gene decreases in response to injury and stays decreased with ephrinA1-Fc [13]. Previous work has shown it to be expressed in genital tubercle vascular endothelia and expression is regulated by HOXA13, a homeobox gene [74]. HOXA13 is also a transcriptional regulator of appropriate vessel morphology and function of the developing placenta [75]. In a recent study, we showed that the EphA6-R localizes to the vasculature and increased in response to injury in EphA2-R-M mouse hearts [76], suggesting that EphA6-R compensates for the absence of EphA2-R. In the current study, only EphA6 was different between uninjured normoglycemic and hyperglycemic WT and EphA2-R-M mouse hearts. While not altered significantly by hyperglycemia at 4 days post-MI in either strain, it was elevated post-MI in all groups. EphA6 is expressed in adult human monocytes and coronary artery endothelial cells [77]. Therefore, given that several ephrins and Eph receptors are upregulated during injury in various cell types, they may play a role in chemotaxis and extravasation and provide insights into the mechanisms of inflammatory diseases and damage resolution [78–80].

Conclusions

This research ratifies our understanding of the negative impact of hyperglycemia on the form and function of the wild type heart. Hyperglycemia leads to decreased survival following MI, impaired compensatory hypertrophy of cardiomyocytes, and increased interstitial fibrosis of the left ventricle. Superimposing hyperglycemia on the infarcted transgenic EphA2-R-M mouse impaired the response to injury and significantly increased mortality. These studies provide the baseline data needed to begin

unraveling the role of EphA2-R in modulating inflammation and evaluate the potential impact of ephrinA1-Fc use in the treatment of MI in the hyperglycemic myocardium.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

AD compiled, analyzed, and interpreted all data and revised the manuscript. SK carried out the mouse breeding, performed the RT-PCR, western blotting, histology and immunohistochemistry, and blood pressure experiments. AC assisted with animal handling and husbandry during these studies, and performed blood glucose measures. JV designed the study, obtained the funding, performed the STZ injections, surgical procedures, and echocardiography, supervised data collection, analyzed and interpreted data, and edited the manuscript to ensure accuracy and inclusion of important intellectual content. All authors have read the manuscript and have given final approval of this version.

Acknowledgements

This work was funded by a pilot grant from the East Carolina Diabetes and Obesity Institute. Jitka Virag is currently funded by a Biotechnology Research Grant from the North Carolina Biotechnology Center and is supported by the Departments of Physiology, and Comparative Medicine, and the East Carolina Diabetes and Obesity Institute. The authors would like to thank Jackson Vuncannon, Jonathan Hodges, Filza Faiz, Tatiana Zuber, Wesley O'Neal, and Mohammed Farooqui for their assistance in collecting the data and helping to prepare this manuscript. We also thank Cameron Schmidt for generating the ephrinA1 model for the image shown on this issue's cover using I-TASSER (v3.0). In addition to the i-Tasser software used, Cameron also use PyMol graphics v1.5.0.1

Author details

¹Department of Physiology, Brody School of Medicine, East Carolina University, 600 Moye Blvd, Greenville, NC 6N-98, USA. ²Department of Comparative Medicine, Brody School of Medicine, East Carolina University, 600 Moye Blvd, Ed Warren Life Sciences Building, Greenville, NC, USA.

Received: 19 March 2014 Accepted: 12 July 2014

Published: 13 August 2014

References

1. Nesto RW, Zarich S: **Acute myocardial infarction in diabetes mellitus: lessons learned from ACE inhibition.** *Circulation* 1998, **97**(1):12–15.
2. Go AS, Mozaffarian D, Roger VL, Benjamin EJ, Berry JD, Borden WB, Bravata DM, Dai S, Ford ES, Fox CS, Franco S, Fullerton HJ, Gillespie C, Hailpern SM, Heit JA, Howard VJ, Huffman MD, Kissela BM, Kittner SJ, Lackland DT, Lichtman JH, Lisabeth LD, Magid D, Marcus GM, Marelli A, Matchar DB, McGuire DK, Mohler ER, Moy CS, Mussolino ME, et al: **Heart Disease and Stroke Statistics—2013 Update: A Report From the American Heart Association.** *Circulation* 2013, **127**(1):e6–e245.
3. Pasquale EB: **Eph-ephrin bidirectional signaling in physiology and disease.** *Cell* 2008, **133**(1):38–52.
4. Kullander K, Klein R: **Mechanisms and functions of Eph and ephrin signalling.** *Nat Rev Mol Cell Biol* 2002, **3**(7):475–486.
5. Brantley DM, Cheng N, Thompson EJ, Lin Q, Brekken RA, Thorpe PE, Muraoka RS, Cerretti DP, Pozzi A, Jackson D, Lin C, Chen J: **Soluble Eph A receptors inhibit tumor angiogenesis and progression in vivo.** *Oncogene* 2002, **21**(46):7011–7026.
6. National Institute of Diabetes and Digestive and Kidney Diseases: **National Diabetes Statistics, 2007 fact sheet.** 2008. 08/28.
7. Brantley-Sieders D, Schmidt S, Parker M, Chen J: **Eph receptor tyrosine kinases in tumor and tumor microenvironment.** *Curr Pharm Des* 2004, **10**(27):3431–3442.
8. Surawska H, Ma PC, Salgia R: **The role of ephrins and Eph receptors in cancer.** *Cytokine Growth Factor Rev* 2004, **15**(6):419–433.
9. Pandey A, Shao H, Marks RM, Polverini PJ, Dixit VM: **Role of B61, the ligand for the Eck receptor tyrosine kinase, in TNF-alpha-induced angiogenesis.** *Science* 1995, **268**(5210):567–569.
10. Cheng N, Brantley DM, Chen J: **The ephrins and Eph receptors in angiogenesis.** *Cytokine Growth Factor Rev* 2002, **13**(1):75–85.

11. Ivanov AI, Romanovsky AA: **Putative dual role of ephrin-Eph receptor interactions in inflammation.** *IUBMB Life* 2006, **58**(7):389–394.
12. Coulthard MG, Morgan M, Woodruff TM, Arumugam TV, Taylor SM, Carpenter TC, Lackmann M, Boyd AW: **Eph/Ephrin signaling in injury and inflammation.** *Am J Pathol* 2012, **181**(5):1493–1503.
13. Dries JL, Kent SD, Virag JA: **Intramyocardial administration of chimeric ephrinA1-Fc promotes tissue salvage following myocardial infarction in mice.** *J Physiol* 2011, **589**(Pt 7):1725–1740.
14. Goichberg P, Bai Y, D'Amaro D, Ferreira-Martins J, Fiorini C, Zheng H, Signore S, Del Monte F, Ottolenghi S, D'Alessandro DA, Michler RE, Hosoda T, Anversa P, Kajstura J, Rota M, Leri A: **The Ephrin A1-EphA2 System Promotes Cardiac Stem Cell Migration After Infarction.** *Circ Res* 2011, **108**(9):1071–1083.
15. O'Neal WT, Griffin WF, Dries-Devlin JL, Kent SD, Chen J, Willis MS, Virag JA: **Ephrin-Eph signaling as a potential therapeutic target for the treatment of myocardial infarction.** *Med Hypotheses* 2013, **80**(6):738–744.
16. Virag JA, Lust RM: **Coronary artery ligation and intramyocardial injection in a murine model of infarction.** *J Vis Exp* 2011, (52):pii: 2581. doi: 10.37971/2581.
17. Yang XP, Liu YH, Rhaleb NE, Kurihara N, Kim HE, Carretero OA: **Echocardiographic assessment of cardiac function in conscious and anesthetized mice.** *Am J Physiol* 1999, **277**(5 Pt 2):H1967–H1974.
18. Beckstead JH: **A simple technique for preservation of fixation-sensitive antigens in paraffin-embedded tissues.** *J Histochem Cytochem* 1994, **42**(8):1127–1134.
19. Ismail JA, Poppa V, Kemper LE, Scatena M, Giachelli CM, Coffin JD, Murry CE: **Immunohistologic labeling of murine endothelium.** *Cardiovasc Pathol* 2003, **12**(2):82–90.
20. Virag JA, Rolle ML, Reece J, Hardouin S, Feigl EO, Murry CE: **Fibroblast growth factor-2 regulates myocardial infarct repair: effects on cell proliferation, scar contraction, and ventricular function.** *Am J Pathol* 2007, **171**(5):1431–1440.
21. Virag JI, Murry CE: **Myofibroblast and endothelial cell proliferation during murine myocardial infarct repair.** *Am J Pathol* 2003, **163**(6):2433–2440.
22. Virag JA, Dries JL, Easton PR, Friesland AM, DeAntonio JH, Chintalgattu V, Cozzi E, Lehmann BD, Ding JM, Lust RM: **Attenuation of myocardial injury in mice with functional deletion of the circadian rhythm gene mPer2.** *Amy Physiol Heart Circ Physiol* 2010, **298**(3):H1088–H1095.
23. Miura T, Miki T: **Limitation of myocardial infarct size in the clinical setting: current status and challenges in translating animal experiments into clinical therapy.** *Basic Res Cardiol* 2008, **103**(6):501–513.
24. Cleutjens JP, Blankesteijn WM, Daemen MJ, Smits JF: **The infarcted myocardium: simply dead tissue, or a lively target for therapeutic interventions.** *Cardiovasc Res* 1999, **44**(2):232–241.
25. Gustafsson AB, Gottlieb RA: **Mechanisms of apoptosis in the heart.** *J Clin Immunol* 2003, **23**(6):447–459.
26. Folkman J: **Angiogenesis research: from laboratory to clinic.** *Forum (Genova)* 1999, **9**(3 Suppl 3):59–62.
27. Kutschka I, Kofidis T, Chen IY, von Degenfeld G, Zwierzchoniewska M, Hoyt G, Arai T, Lebl DR, Hendry SL, Sheikh AY, Cooke DT, Connolly A, Blau HM, Gambhir SS, Robbins RC: **Adenoviral human BCL-2 transgene expression attenuates early donor cell death after cardiomyoblast transplantation into ischemic rat hearts.** *Circulation* 2006, **114**(1 Suppl):1174–1180.
28. Yau TM, Li G, Weisel RD, Reheman A, Jia ZQ, Mickle DA, Li RK: **Vascular endothelial growth factor transgene expression in cell-transplanted hearts.** *J Thorac Cardiovasc Surg* 2004, **127**(4):1180–1187.
29. Hwang H, Kloner RA: **Improving regenerating potential of the heart after myocardial infarction: factor-based approach.** *Life Sci* 2010, **86**(13–14):461–472.
30. Dorn GW 2nd, Diwan A: **The rationale for cardiomyocyte resuscitation in myocardial salvage.** *J Mol Med (Berl)* 2008, **86**(10):1085–1095.
31. Urbich C, Rossig L, Dimmeler S: **Restoration of cardiac function with progenitor cells.** *Novartis Found Symp* 2006, **274**:214–223. discussion 223–217, 272–216.
32. Laflamme MA, Zbinden S, Epstein SE, Murry CE: **Cell-based therapy for myocardial ischemia and infarction: pathophysiological mechanisms.** *Annu Rev Pathol* 2007, **2**:307–339.
33. Bartunek J, Vanderheyden M, Hill J, Terzic A: **Cells as biologics for cardiac repair in ischaemic heart failure.** *Heart* 2010, **96**(10):792–800.
34. Brownlee M: **The pathobiology of diabetic complications: a unifying mechanism.** *Diabetes* 2005, **54**(6):1615–1625.
35. Bugger H, Abel ED: **Mitochondria in the diabetic heart.** *Cardiovasc Res* 2010, **88**(2):229–240.
36. Yan SF, Ramasamy R, Schmidt AM: **The receptor for advanced glycation endproducts (RAGE) and cardiovascular disease.** *Expert Rev Mol Med* 2009, **11**:e9.
37. Li Z, Zhang T, Dai H, Liu G, Wang H, Sun Y, Zhang Y, Ge Z: **Involvement of endoplasmic reticulum stress in myocardial apoptosis of streptozocin-induced diabetic rats.** *J Clin Biochem Nutr* 2007, **41**(1):58–67.
38. Kuethle F, Sigusch HH, Bornstein SR, Hilbig K, Kamvissi V, Figulla HR: **Apoptosis in patients with dilated cardiomyopathy and diabetes: a feature of diabetic cardiomyopathy?** *Horm Metab Res* 2007, **39**(9):672–676.
39. Tian XF, Cui MX, Yang SW, Zhou YJ, Hu DY: **Cell death, dysglycemia and myocardial infarction.** *Biomedical Reports* 2013, **1**(3):341–346.
40. Yang SW, Zhou YJ, Tian XF, Pan GZ, Liu YY, Zhang J, Guo ZF, Chen SY, Gao ST, Du J, Jia DA, Fang Z, Hu B, Han HY, Gao F, Hu DY, Xu YY, Beijing H, Metabolism Survey Study G: **Association of dysglycemia and all-cause mortality across the spectrum of coronary artery disease.** *Mayo Clin Proc* 2013, **88**(9):930–941.
41. Lonborg J, Vejstrup N, Kelbaek H, Nepper-Christensen L, Jorgensen E, Helqvist S, Holmvang L, Saunamaki K, Botker HE, Kim WY, Clemmensen P, Treiman M, Engstrom T: **Impact of Acute Hyperglycemia on Myocardial Infarct Size, Area at Risk, and Salvage in Patients With STEMI and the Association With Exenatide Treatment: Results From a Randomized Study.** *Diabetes* 2014, **63**(7):2474–2485.
42. Marfella R, Siniscalchi M, Esposito K, Sellitto A, De Fanis U, Romano C, Portoghese M, Siciliano S, Nappo F, Sasso FC, Mininni N, Ciacciapuoti F, Lucivero G, Giunta R, Verza M, Giugliano D: **Effects of stress hyperglycemia on acute myocardial infarction: role of inflammatory immune process in functional cardiac outcome.** *Diabetes Care* 2003, **26**(11):3129–3135.
43. Eguchi M, Kim YH, Kang KW, Shim CY, Jang Y, Dorval T, Kim KJ, Sweeney G: **Ischemia-reperfusion injury leads to distinct temporal cardiac remodeling in normal versus diabetic mice.** *PLoS One* 2012, **7**(2):e30450.
44. Connor RJ, Menzel P, Pasquale EB: **Expression and tyrosine phosphorylation of Eph receptors suggest multiple mechanisms in patterning of the visual system.** *Dev Biol* 1998, **193**(1):21–35.
45. Ogawa K, Pasqualini R, Lindberg RA, Kain R, Freeman AL, Pasquale EB: **The ephrin-A1 ligand and its receptor, EphA2, are expressed during tumor neovascularization.** *Oncogene* 2000, **19**(52):6043–6052.
46. Bovenkamp DE, Greer PA: **Degenerate PCR-based cloning method for Eph receptors and analysis of their expression in the developing murine central nervous system and vasculature.** *DNA Cell Biol* 2001, **20**(4):203–213.
47. Nakamoto M, Bergemann AD: **Diverse roles for the Eph family of receptor tyrosine kinases in carcinogenesis.** *Microsc Res Tech* 2002, **59**(1):58–67.
48. Duxbury MS, Ito H, Zinner MJ, Ashley SW, Whang EE: **EphA2: a determinant of malignant cellular behavior and a potential therapeutic target in pancreatic adenocarcinoma.** *Oncogene* 2004, **23**(7):1448–1456.
49. Lin YG, Han LY, Kamat AA, Merritt WM, Landen CN, Deavers MT, Fletcher MS, Urbauer DL, Kinch MS, Sood AK: **EphA2 overexpression is associated with angiogenesis in ovarian cancer.** *Cancer* 2007, **109**(2):332–340.
50. Wykosky J, Debinski W: **The EphA2 receptor and ephrinA1 ligand in solid tumors: function and therapeutic targeting.** *Mol Cancer Res* 2008, **6**(12):1795–1806.
51. Okazaki T, Ni A, Baluk P, Ayeni OA, Kearley J, Coyle AJ, Humbles A, McDonald DM: **Capillary defects and exaggerated inflammatory response in the airways of EphA2-deficient mice.** *Am J Pathol* 2009, **174**(6):2388–2399.
52. Funk SD, Yurdagul A Jr, Albert P, Traylor JG Jr, Jin L, Chen J, Orr AW: **EphA2 Activation Promotes the Endothelial Cell Inflammatory Response: A Potential Role in Atherosclerosis.** *Arterioscler Thromb Vasc Biol* 2012, **32**(3):686–695.
53. Funk SD, Orr AW: **Ephs and ephrins resurface in inflammation, immunity, and atherosclerosis.** *Pharmacol Res* 2013, **67**(1):42–52.
54. Carpenter TC, Schroeder W, Stenmark KR, Schmidt EP: **Eph-A2 promotes permeability and inflammatory responses to bleomycin-induced lung injury.** *Am J Respir Cell Mol Biol* 2012, **46**(1):40–47.
55. Zhou N, Zhao WD, Liu DX, Liang Y, Fang WG, Li B, Chen YH: **Inactivation of EphA2 promotes tight junction formation and impairs angiogenesis in brain endothelial cells.** *Microvasc Res* 2011, **82**(2):113–121.
56. Thundiyil J, Manzanero S, Pavlovski D, Cully TR, Lok KZ, Widiapradja A, Chunduri P, Jo DG, Naruse C, Asano M, Lounikou BS, Sobey CG, Coulthard MG,

- Arumugam TV: Evidence that the EphA2 receptor exacerbates ischemic brain injury. *PLoS One* 2013, **8**(1):e53528.
57. Baldwin C, Chen ZW, Bedirian A, Yokota N, Nasr SH, Rabb H, Lemay S: **Upregulation of EphA2 during in vivo and in vitro renal ischemia-reperfusion injury: role of Src kinases.** *Am J Physiol Renal Physiol* 2006, **291**(5):F960–F971.
58. Jellinghaus S, Poitz DM, Ende G, Augstein A, Weinert S, Stutz B, Braun-Dullaeus RC, Pasquale EB, Strasser RH: **Ephrin-A1/EphA4-mediated adhesion of monocytes to endothelial cells.** *Biochim Biophys Acta* 2013, **1833**(10):2201–2211.
59. Chan B, Sukhatme VP: **Receptor tyrosine kinase EphA2 mediates thrombin-induced upregulation of ICAM-1 in endothelial cells in vitro.** *Thromb Res* 2009, **123**(5):745–752.
60. Dixit VM, Green S, Sarma V, Holzman LB, Wolf FW, O'Rourke K, Ward PA, Prochownik EV, Marks RM: **Tumor necrosis factor-alpha induction of novel gene products in human endothelial cells including a macrophage-specific chemotaxin.** *J Biol Chem* 1990, **265**(5):2973–2978.
61. Holzman LB, Marks RM, Dixit VM: **A novel immediate-early response gene of endothelium is induced by cytokines and encodes a secreted protein.** *Mol Cell Biol* 1990, **10**(11):5830–5838.
62. Ivanov AI, Steiner AA, Scheck AC, Romanovsky AA: **Expression of Eph receptors and their ligands, ephrins, during lipopolysaccharide fever in rats.** *Physiol Genomics* 2005, **21**(2):152–160.
63. Kolattukudy PE, Niu J: **Inflammation, endoplasmic reticulum stress, autophagy, and the monocyte chemoattractant protein-1/CCR2 pathway.** *Circ Res* 2012, **110**(1):174–189.
64. Ren G, Dewald O, Frangogiannis NG: **Inflammatory mechanisms in myocardial infarction.** *Current Drug Targets Inflammation Allergy* 2003, **2**(3):242–256.
65. Carter AM: **Complement activation: an emerging player in the pathogenesis of cardiovascular disease.** *Scientifica* 2012, **2012**:402783.
66. Busche MN, Stahl GL: **Role of the complement components C5 and C3a in a mouse model of myocardial ischemia and reperfusion injury.** *German Med Sci* 2010, **8**:doc20.
67. Goldbergova MP, Parenica J, Jarkovsky J, Kala P, Poloczek M, Manousek J, Kluz K, Kubkova L, Littnerova S, Tesak M, Toman O, Pavek N, Cermakova Z, Tomandl J, Vasku A, Spinar J: **The association between levels of tissue inhibitor of metalloproteinase-1 with acute heart failure and left ventricular dysfunction in patients with ST elevation myocardial infarction treated by primary percutaneous coronary intervention.** *Gen Test Mol Biomark* 2012, **16**(10):1172–1178.
68. Abohashem-Aly AA, Meng X, Li J, Sadaria MR, Ao L, Wennergren J, Fullerton DA, Raeburn CD: **DITPA, a thyroid hormone analog, reduces infarct size and attenuates the inflammatory response following myocardial ischemia.** *J Surg Res* 2011, **171**(2):379–385.
69. Pixley FJ, Stanley ER: **CSF-1 regulation of the wandering macrophage: complexity in action.** *Trends Cell Biol* 2004, **14**(11):628–638.
70. Frangogiannis NG, Mendoza LH, Ren G, Akrivakis S, Jackson PL, Michael LH, Smith CW, Entman ML: **MCSF expression is induced in healing myocardial infarcts and may regulate monocyte and endothelial cell phenotype.** *Am J Physiol Heart Circ Physiol* 2003, **285**(2):H483–H492.
71. Iyigun T, Teskin O, Enc Y, Cakmak M, Iyigun M, Keser S, Camur G, Komurcu G, Sargin M, Ozay B, Dagsali S: **Does the level of soluble intercellular adhesion molecule 1 predict myocardial injury before cardiac markers increase?** *Heart Surg Forum* 2008, **11**(6):E352–E356.
72. Shai I, Pischon T, Hu FB, Ascherio A, Rifai N, Rimm EB: **Soluble intercellular adhesion molecules, soluble vascular cell adhesion molecules, and risk of coronary heart disease.** *Obesity* 2006, **14**(11):2099–2106.
73. Jehle J, Staudacher I, Wiedmann F, Schweizer P, Becker R, Katus H, Thomas D: **Regulation of apoptosis in HL-1 cardiomyocytes by phosphorylation of the receptor tyrosine kinase EphA2 and protection by lithocholic acid.** *Br J Pharmacol* 2012, **167**(7):1563–1572.
74. Shaut CA, Saneyoshi C, Morgan EA, Knosp WM, Sexton DR, Stadler HS: **HOXA13 directly regulates EphA6 and EphA7 expression in the genital tubercle vascular endothelia.** *Dev Dyn* 2007, **236**(4):951–960.
75. Shaut CA, Keene DR, Sorensen LK, Li DY, Stadler HS: **HOXA13 is essential for placental vascular patterning and labyrinth endothelial specification.** *PLoS Genet* 2008, **4**(5):e1000073.
76. O'Neal WT: **Deletion of the EphA2 receptor exacerbates myocardial injury and the progression of ischemic cardiomyopathy.** *Front Physiol* 2014, **5**:article 132.
77. Sakamoto A, Sugamoto Y, Tokunaga Y, Yoshimuta T, Hayashi K, Konno T, Kawashiri MA, Takeda Y, Yamagishi M: **Expression profiling of the ephrin (EFN) and Eph receptor (EPH) family of genes in atherosclerosis-related human cells.** *J Int Med Res* 2011, **39**(2):522–527.
78. Yamagishi M, Higashikata T, Ishibashi-Ueda H, Sasaki H, Ogino H, Iihara K, Miyamoto S, Nagaya N, Tomoike H, Sakamoto A: **Sustained upregulation of inflammatory chemokine and its receptor in aneurysmal and occlusive atherosclerotic disease: results from tissue analysis with cDNA microarray and real-time reverse transcriptional polymerase chain reaction methods.** *Circ J* 2005, **69**(12):1490–1495.
79. O'Donnell R, Breen D, Wilson S, Djukanovic R: **Inflammatory cells in the airways in COPD.** *Thorax* 2006, **61**(5):448–454.
80. Bouloumie A, Curat CA, Sengenès C, Lolmede K, Miranville A, Busse R: **Role of macrophage tissue infiltration in metabolic diseases.** *Curr Opin Clin Nutr Metab Care* 2005, **8**(4):347–354.

doi:10.1186/s12933-014-0114-y

Cite this article as: DuSablón et al.: EphA2-receptor deficiency exacerbates myocardial infarction and reduces survival in hyperglycemic mice. *Cardiovascular Diabetology* 2014 **13**:114.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

