Abstract

Multi-walled carbon nanotubes (MWCNTs) are increasingly used in industry and in nanomedicine raising safety concerns, especially during unique life-stages such as pregnancy. We hypothesized that MWCNT exposure during pregnancy will increase vascular tissue contractile responses by increasing Rho kinase signaling. Pregnant (17–19 gestational days) and non-pregnant Sprague Dawley rats were exposed to 100 μg/kg of MWCNTs by intratracheal instillation or intravenous administration. Vasoactive responses of uterine, mesenteric, aortic and umbilical vessels were studied 24 hours post-exposure by wire myography. The contractile responses of the vessel segments were different between the pregnant and non-pregnant rats, following MWCNT exposure. Maximum stress generation in the uterine artery segments from the pregnant rats following pulmonary MWCNT exposure was increased in response to angiotensin II by 4.9 mN/mm² (+118%), as compared to the naïve response and by 2.6 mN/mm² (+40.7%) as compared to the vehicle exposed group. Following MWCNT exposure, serotonin induced approximately 4 mN/mm² increase in stress generation of the mesenteric artery from both pregnant and non-pregnant rats as compared to the vehicle response. A significant contribution of the dispersion medium was identified as inducing changes in the contractile properties following both pulmonary and intravenous exposure to MWCNTs. Wire myographic studies in the presence of a Rho kinase inhibitor and RhoA and Rho kinase mRNA/protein expression of rat aortic endothelial cells were unaltered following exposure to MWCNTs, suggesting absent/minimal contribution of Rho kinase to the enhanced contractile responses following MWCNT exposure. The reactivity of the umbilical vein was not changed; however, mean fetal weight gain was reduced with dispersion media and MWCNT exposure by both routes. These results suggest a susceptibility of the vasculature during gestation to MWCNT and their dispersion media-induced vasoconstriction, predisposing reduced fetal growth during pregnancy.

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Vascular tissue contractility; Pregnancy; Uterine artery; Umbilical vein; Nanotoxicology; MWCNTs

Introduction
An increasing number of single- and multi-walled carbon nanotubes (SWCNTs and MWCNTs) are being designed and produced for various industrial and biomedical applications such as tracers of malignant cells, immunomodulators, contrast agents and as scaffolds in tissue engineering [1,2]. Pulmonary exposure to MWCNTs are reported to be associated with adverse effects similar to asbestos exposure [3] involving impairment in pulmonary functions [4] and activation of inflammatory responses in mesothelial cells [5]. MWCNTs are known to be taken-up by bronchial epithelial cells, increase pro-inflammatory cytokine production and induce cytotoxicity in in vitro studies [6,7]. When considering their bio-distribution, MWCNTs translocate to the lymph nodes following intratracheal instillation [8,9] and potentially to other extra-pulmonary organs including the liver, kidney and heart and contributing to various toxico-pathologies [10]. The extra-pulmonary effects of MWCNT exposure is reported to be associated with impairment of endothelium dependent relaxation in coronary arterioles [11] and increased coronary vascular tone enhancing indices of ischemia reperfusion injury [12]. The adverse pulmonary effects following occupational exposure to carbon nanotubes have been studied extensively in non-pregnant animal models [9,13]. The consequences of MWCNT exposure on the peripheral vascular system are yet to be studied adequately, particularly in the unique physiological stage of pregnancy.

In general, exposure to MWCNTs occurs by inhalation during occupational exposures in industry or in research laboratories [13–15]. Potential biomedical applications could also expose an individual to MWCNTs primarily by the intravenous route [16]. An animal model study on MWCNT exposure during pregnancy reported minimal effects on fetal development and maternal well-being following oral exposure to 8–1000 mg/kg/day of MWCNTs [17]. The expansive vascular remodeling that takes place during pregnancy [18,19] may predispose the maternal and fetal vasculature to be sensitive to nanomaterial exposures by various routes (i.e. pulmonary and intravenous) where increased concentrations of MWCNTs may directly reach the circulation. The consequence of any changes in vascular reactivity can potentially negatively influence the placental blood supply, impacting fetal growth and development. Following acute intravenous exposure, pristine carbon nanotubes are redistributed to the reticulo-endothelial system [16,20] with a significant proportion remaining in blood [21]. This is in contrast to functionalized forms, which are reported to be excreted unchanged via the kidney [22,23]. It can be assumed that these nanotubes come in direct contact with the vascular endothelium during their circulation and these interactions can potentially induce changes in vascular reactivity during pregnancy by various mechanisms.
Multiple vasoconstrictor agents including phenylephrine, endothelin 1, angiotensin II and serotonin act through \( G_q \) protein coupled receptors to regulate smooth muscle contraction in the vasculature. Downstream of this receptor, the RhoA/Rho kinase pathway plays a critical role in mediating contractile response in vascular smooth muscle cells. The active form of RhoA promotes activation of the Rho kinase (ROCK) that inhibits MLC phosphatase (MLCP) activity, the dephosphorylation of myosin and subsequent relaxation [24]. Alterations in the RhoA/Rho kinase pathway is reported to be involved in endothelial dysfunction, inflammation [25,26] and with exposure to particulate matter [27].

We hypothesized that MWCNT exposure during pregnancy would increase the contractile responses in uterine and placenta derived blood vessels by increasing the RhoA/Rho kinase activity. We also hypothesized that there will be differential effects on the contractile responses dependent on the route of exposure and the vascular bed location. Intratracheal instillation and intravenous administration were used as the two routes of exposure to identify these differential effects within thoracic aorta, mesenteric and uterine arterial segments.

**Methods and Materials**

**MWCNT suspensions for exposure**

Multi-walled carbon nanotubes (MWCNTs) were a generous gift from NanoTechLabs Inc. (Yadkinville, NC, USA) and the dry powder form was previously characterized [4]. The commercial grade, non-functionalized, hydrophobic carbon based nanotubes were suspended in non-polar solvents/dispersal media prior to *in vivo* exposure. MWCNTs for intratracheal instillation was suspended in 10% clinical grade surfactant (Infasurf\textsuperscript{®}, ONY, Inc., Amherst, NY, USA), to mimic protein-lipid coating from lung surfactant, in sterile 0.9% saline (0.9% NaCl, B. Braun Medical Inc., CA, USA) as previously described [4] to a concentration of 150 \( \mu \)g/ml and the mixture was cup-horn sonicated for 2 minutes at 65% amplitude for a total energy of 10,817 Joules, using a Misonix ultrasonic liquid processor -1510R-MTH (Branson Ultrasonics Corp. Danbury, CT, USA). This suspension will be referred to as “(S)-MWCNTs” and had been previously characterized by Wang et al. [4]. Additionally, the MWCNTs were suspended in the dispersion media modified from Bihari et al. [28] for intravenous administration to coat with a vascular compartment protein. Briefly, this dispersion media contained 0.6mg/ml rat serum albumin (Sigma, A6272), 0.01 mg/ml 1, 2-dipalmitoyl-sn-glycero-3-phosphocholine (DPPC, Sigma F-0763) in phosphate buffered saline (Sigma D5652 1X) and sonicated using the probe sonicator at 40% amplitude for 15 seconds. This dispersion medium will be referred to as “DPPC/RSA”. A MWCNT suspension of 150 \( \mu \)g/ml was made with DPPC/RSA and the mixture was cup-horn sonicated using a Misonix ultrasonic liquid processor -1510R-MTH (Branson Ultrasonics Corp. Danbury, CT, USA) at 65% amplitude for 2 minutes. This intravenous suspension will be referred to as “(D)-MWCNTs” and was previously described by Wang et al. [29].

**Sprague Dawley rats**

Pregnant and non-pregnant female, 10–12 week old Sprague Dawley rats were purchased from Charles River Laboratories (USA). All rats were acclimated for one week in East
Carolina University (ECU) Department of Comparative Medicine’s animal facility, housed under 12 hour light/dark cycles with standard rat chow and water provided *ad libitum*. The pregnant rat arrived in the facility between 9–12 days of gestation and the body weight was monitored once in every three days to assess the progression of pregnancy. All animal handling and exposure procedures were approved by the ECU Institutional Animal Care and Use Committee.

**MWCNT exposure and dosing**

Each pregnant and non-pregnant female rat was randomly assigned to either the MWCNT exposure or dispersion medium control group for each route of exposure to include a minimum of six animals in a group. Matched gestational day pregnancies were used to compare vehicle vs. MWCNT effects. The pregnant rats were exposed between 17–19 days of gestation, compatible with the third trimester of human pregnancy (i.e. late gestational stage). Rats were anesthetized using 2–3% isoflurane (Webster Veterinary, USA) dispersed in oxygen for exposure procedures. The 150 μg/ml MWCNT suspension was administered as a mass based dose of 100 μg/kg by weighing each rat just before the exposure. (S)-MWCNTs suspension or 10% surfactant was instilled intratracheally (IT) as previously described \[4,12\] for pulmonary exposure. A group of non-pregnant female rats was exposed to IT (S)-MWCNTs or 10% surfactant to evaluate any effect of life stage (pregnant vs. non-pregnant) on vascular tissue contractility. Intravenous (IV) administration of 100 μg/kg (D)-MWCNTs or DPPC/RSA was done in the pregnant rats through the tail vein using a 25G needle. Ten to twelve weeks old, pregnant (GD 17–19) and non-pregnant female rats were used as naïve controls.

**Tissue and sample collection**

All rats were anesthetized in a transparent sealed receptacle containing gauze soaked with 70% isoflurane (Webster Veterinary, USA) in propylene glycol (Amersco, OH, USA), separated from the animal by a desiccator plate/grid prior to euthanasia. Twenty-four hours following administration of the MWCNTs or vehicle, the rats were subjected to a midline incision and euthanized by pneumothorax. Whole blood (~1 ml) was withdrawn directly from the maternal right ventricle. A pooled fetal blood sample was collected from three fetuses in each pregnant dam (blood from these three fetuses were considered as one sample). Maternal and fetal whole blood samples were centrifuged (20,400 × g for 20 minutes), and serum supernatant was stored at −80°C for cytokine analysis.

**Isolation of vessel segments**

Three arterial beds and the umbilical vein were selected for pharmacological myographic studies. The uterine and mesenteric vascular beds were selected as they manifest both structural and functional changes during pregnancy [30,31] with the uterine vasculature undergoing significant remodeling [32]. The thoracic aorta was included as proximal conduit vessel not anticipated to undergo significant remodeling, but still may express changes in pharmacological responses. Both uterine horns with the vascular arcades, small intestinal loop with superior mesenteric arcade and thoracic aorta were carefully excised and placed in ice cold physiological saline solution (PSS; mM) 140 NaCl, 5.0 KCl, 1.6 CaCl₂, 1.2 MgSO₄, 1.2 MOPS (3-[N-morpholino]-propane sulfonic acid), 6 D-glucose, 0.02 EDTA, and a pH of
Arterial segments with a length of 0.5 – 2.0 mm were isolated from the mid region of the main uterine artery (diameter 150–300 μm), first order mesenteric artery (diameter 150–250 μm), and thoracic aorta (diameter 2–3 mm). Two umbilical vein segments (diameter 400–550 μm) from umbilical cords of different fetuses implanted in the mid-uterine region were isolated from each dam.

Maternal and fetal serum cytokine analysis

The targets for maternal and fetal serum cytokine analysis were selected based on the reported cytokine targets in previous MWCNT exposure studies [4,33,34]. The selected serum cytokines and chemokines (IL6, IL10, TNFα, MCP1, VEGF, INFγ, and IL1β) were assessed using Milliplex MAP Cytokine/Chemokine Panel and Immunoassay (EMD Millipore MA, USA) according to the manufacturer’s directions. Assays were run using Luminex 100/200 (Luminex, Austin, TX) and results reported using Luminex xPONENT® software versions 2.3/3.1.

Bronchoalveolar lavage cytology

Twenty-four hours following exposure to MWCNTs or dispersion media, a bronchoalveolar lavage (BAL) was performed on adult female rats as described previously [4]. Briefly, the right lung was lavaged in situ three times with repeated flushes of 26.25 mL/kg body weight of ice-cold Hanks balanced salt solution. The BAL fluids were centrifuged and the total number of cells was calculated using an automated cell counter (Cellometer, Nexcelom Bioscience, and Lawrence, MA, USA). A sample of 20,000 cells was centrifuged using a Cytospin IV (Shandon Scientific Ltd., Cheshire, UK) and stained with a three-step hematology stain (Richard Allan Scientific, Kalamazoo, MI, USA). The differential cell count was determined by morphology, evaluating 300 cells per slide using light microscopy and each cell count is reported as a percentage of 20,000 cells.

Wire myographic studies

The dissected vessel segments were mounted into a DMT 610M multi-channel wire myograph system (Danish Myo Technology, Aarhus N, Denmark) using 40 μm wires or pins. All vessel segments were bathed in PSS at 37°C, bubbled with medical grade air during the entire myographic studies. The optimal resting tension for each arterial segment was established at 90% of internal circumference (IC) produced at tensions equivalent to 100 mmHg (13.3 kPa). A depolarization response with K⁺PSS (109 mM K⁺ equal molar substitution of Na⁺) was used to assess the vessel viability and segments that developed a stress response of greater than 1 mN/mm² were considered viable. Endothelial viability was assessed by adding 3.0 μM acetylcholine during a 1 μM phenylephrine pre-contraction. Each segment along the arterial tree adapts to different hemodynamic conditions including blood pressure and autonomic innervations. Such adaptations will express different receptor classes and relative amounts of their subtypes, thus it is necessary to investigate different agonist responses in different vascular segments. All three arterial (uterine, mesenteric and aortic) vessel segments were subjected to cumulative concentrations of phenylephrine (0.001–30 μM), endothelin-1 (0.0001–1 μM) and acetylcholine (0.0001–30 μM). Angiotensin II (0.0001–0.1 μM) and serotonin (0.001–1 μM) was used to study the uterine and mesenteric arteries respectively. The force generated by each vessel segment at each
concentration was recorded using Lab Chart (ADI Instruments, CO, USA). The force was then normalized to the surface area of the vessel to determine the active stress generated in response to different agonists.

The umbilical vein segments were stretched and set to an IC equal to 90% of the IC when the wall tension is equivalent to 20 mmHg (5.1 kPa) [35]. The viability was assessed using K+PSS. The segments were pre-contracted with thromboxane-mimetic U46619 (1 μM) and subjected to cumulative concentrations of acetylcholine (0.0001–30 μM), followed by 1 μM sodium nitroprusside.

**Cell culture, mRNA and protein quantification**

Rat aortic endothelial cells (RAEC) in the *in vitro* studies were used to identify the contribution of the endothelial RhoA/Rho kinase signaling following direct exposure to MWCNTs suspended in different media. RAEC were purchased from Cascade Biologics (Eugene, OR, USA), grown with Dulbecco’s Modified Eagle Medium (DMEM) and cultured at >90% confluence were treated with (S)-MWCNTs or (D)-MWCNTs over a 1–10 μg/cm² dose range for 2–12 hours. Untreated cells, 10% surfactant treated cells and DPPC/RSA treated cells were used as controls. Real time-PCR was done as described previously [24] to identify any changes Rhoa, ROCK1, ROCK2 and eNOS mRNA expression following 2 hours of exposure to MWCNTs. Following 12-hour *in vitro* exposure to MWCNTs or dispersion medium, In-Cell Western Assay (Li-Cor Biosciences, Lincoln, NE, USA) was performed to assess the changes in target protein expression [25] for Rhoa, ROCK1, ROCK2 and eNOS. The cell media was removed and the cells were immediately fixed with 3% formaldehyde followed by permeabilization with 0.1% Triton-X, blocked with Odyssey blocking buffer (LI-COR Biosciences, Lincoln, NE, USA). The primary antibodies for Rhoa (1:1000), ROCK1 (1:500), ROCK2 (1:500) and eNOS (1:1000) (Santa Cruz Biotechnology Inc., USA and Cell Signaling Danvers, MA, USA) were added and plate incubated overnight. IRDye 800CW Secondary Antibodies (LI-COR Biosciences, Lincoln, NE, USA) were used in 1:10000 dilutions to identify target proteins. The DNA was stained with DRAQ5 and Sapphire 700 (Cell Signaling, Danvers, MA, USA) for cell number normalization. The Fluorescence was detected, quantified and analyzed using Li-Cor Odyssey Infrared Imaging System and software (LI-COR Biosciences, Lincoln, NE, USA).

**Measurement of the fetal and placental weight**

Body weights of pregnant dams were recorded just before sacrifice and the litter size was recorded before uterine vessel isolation. Three fetuses were isolated from each dam from the mid-uterine region and individual weights were measured using Ohaus Explorer Analytical Balance (Ohaus Corporation, NJ, USA). The blot weights of the placentae attached to the same fetuses were also recorded. The individual weights of the fetuses/placentae were then grouped according to the day of gestation at sacrifice and their mean weight was used for comparison between the treatment groups. Each day of gestation included the pups from at least 2 dams.

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Statistical analysis

Statistical analysis was done utilizing GraphPad Prism 5 software (San Diego, CA) and data is presented as mean ± SEM (standard error of mean). Repeated measures analysis of variance [36] and Bonferroni post hoc test were used to compare the concentration-responses of different agonists and the differences were considered statistically significant if \( p<0.05 \). In addition, each concentration-response curve was also compared across treatment groups using a regression analysis by examining the best-fit values [36]. EC\(_{50}\) values for concentration-responses in myographic studies were determined using the Hill equation. A two tailed t test was used compare mean EC\(_{50}\), umbilical vein stress generation, and cytokine expression levels between different treatment and control groups. One way ANOVA and Turkey post-hoc test was used for the analysis of fetal and placental weight on each day of gestation.

Results

Characterization of MWCNT suspensions

The MWCNT suspension in 10% surfactant in saline [(S)-MWCNTs] has been previously characterized by Wang et al. [4]. MWCNTs used in this study had a mean diameter of 22.5 ± 1.3 nm with a bimodal distribution with peaks at 12.5 and 25 nm and a length range of 10–100 μm and a surface area of 113.1 m\(^2\)/g). The zeta potential of the particles in (S)-MWCNTs suspension was −57.3 mV with a mean hydrodynamic size of 915 nm. MWCNTs suspended in the DPPC, serum albumin and sterile phosphate buffered saline medium [(D)-MWCNTs] was characterized previously by Wang et al. [29] with a zeta potential of −20.8 mV with a mean hydrodynamic size of 793 nm.

Maternal serum cytokine analysis

The mean values serum cytokine levels of pregnant and non-pregnant rats 24 hours following exposure to MWCNTs or dispersion media for each route of exposure are reported in Table 1. The baseline cytokine levels in the naïve rats were relatively higher in the non-pregnant group compared to the pregnant group for all cytokines assessed except VEGF. In general, the cytokine profiles for the rats exposed to vehicle or MWCNT were lower in the non-pregnant group while a few changes were noted in the pregnant group. IL1β level was increased by five fold in the pregnant group (when compared to the naive) following IV DPPC/RSA. TNFα levels were increased more than six fold in the serum following exposure to both dispersion media (10% surfactant and DPPC/RSA) and increased with IV (D)-MWCNTs.

Maternal Bronchoalveolar Lavage (BAL) cell counts

The percentages of differential cell counts in the bronchoalveolar lavage fluid are graphed in Supplementary Figure 1 and MWCNT induced changes in these cell counts were observed only in the pregnant group. The mean percentage of macrophages was 4.2% lower in the naïve pregnant group compared to the naïve non-pregnant group, and increased during pregnancy following exposure to both vehicles (by 4.8% with 10% surfactant and by 5.0% with DPPC/RSA) and intravenous (D)-MWCNT exposure (by 4.3%) compared to the naive.
In contrast, the mean epithelial cell percentage was 4% higher in the naïve pregnant group compared to the naïve non-pregnant group and was reduced by ~5% during pregnancy by following exposure to both vehicles and MWCNTs by both routes of exposure when compared to the naïve. The percentage of neutrophils were highest following exposure to (S)-MWCNTs via intratracheal instillation but was less than 1% of the total BAL cell counts. The percentages of eosinophils were not significantly different following exposure to MWCNTs during pregnancy.

Responses of arterial segments 24 hours post-exposure to intratracheal (IT) instillation of (S)-MWCNTs or 10% surfactant in pregnant and non-pregnant female rats

The contractile responses of the vessel segments from the pregnant and non-pregnant rats were different following IT exposure to (S)-MWCNTs. In general, the pregnant group manifested increased contractile responses in multiple vascular beds that were in part contributed to by the dispersal media as opposed to minimal changes in observed in the non-pregnant group.

Main uterine artery

The main uterine artery segments from pregnant and non-pregnant rats responded differently to the same dose of IT instilled (S)-MWCNTs. The maximum stress generation was increased in the pregnant group in response to phenylephrine by 2.6 mN/mm² (+37%) following IT exposure to (S)-MWCNTs when compared to the naïve, but was not significantly increased when compared to the responses from 10% surfactant group (Figure 1A). The response to angiotensin II following IT exposure to (S)-MWCNT during pregnancy was increased by 4.9 mN/mm² (+118%), as compared to the naïve and by 2.6 mN/mm² (+40.7%) as compared to the 10% surfactant exposed group (Figure 1B). In contrast, the stress generation in response to all 3 agonists was diminished in uterine artery segments from non-pregnant animals following (S)-MWCNTs exposure (Figure 1B, D and F). The relaxation responses to acetylcholine during 30 μM phenylephrine pre-contraction were not different in naive, 10% surfactant and (S)-MWCNTs exposed pregnant groups (Figure 1G), but was diminished ~ 10% following (S)-MWCNTs exposure in the non-pregnant group (Figure 1H). The calculated EC₅₀ values for phenylephrine, angiotensin II, acetylcholine and HA-1077 were not different between the naive, 10% surfactant and (S)-MWCNTs treatment groups. Following (S)-MWCNTs exposure in pregnant rats, calculated EC₅₀ value for endothelin 1 (1.1 ± 0.3 nM) was significantly lower than the naive (3.4 ± 0.6 nM), but not different form the 10% surfactant exposed group (2.2 ± 1.0 nM, Supplementary Table 1).

First order mesenteric artery

The mesenteric artery segments from both pregnant and non-pregnant rats responded in a similar manner following IT (S)-MWCNT exposure. The stress generations in response to serotonin in the first order mesenteric artery segments were increased by ~ 4 mN/mm² following IT (S)-MWCNTs exposure compared to 10% surfactant exposed group (Figure 2E and F). The contractile responses to phenylephrine or endothelin 1 and the relaxation response to acetylcholine were not changed following IT (S)-MWCNT exposure in the pregnant group (Figure 2A, C and G).
The mesenteric artery contractile responses to all 3 agonists were diminished in the non-pregnant rats exposed to 10% surfactant (Figure 2B, D and F), along with an impairment of acetylcholine dependent relaxation response (Figure 2H). Similar to the reported uterine vessels responses, the EC$_{50}$ values of the mesenteric arteries responses following (S)-MWCNT exposure were not different except for endothelin 1 (Supplementary Table 2). The EC$_{50}$ for endothelin 1-mediated responses was decreased in the 10% surfactant (1.4 ± 0.4 nM) group when compared to both naïve (5.4 ± 1.3 nM) and (S)-MWCNTs exposed group (5.0 ± 1.0 nM).

**Thoracic aorta**

The thoracic aortic segments from pregnant and non-pregnant rats responded differently to the IT exposure to (S)-MWCNTs. The contractile response to phenylephrine (0.001–10 μM) was reduced by 0.68 mN/mm$^2$ (25.4%) in the pregnant group following (S)-MWCNT exposure compared to the naïve, but was not different when compared to the 10% surfactant exposed group (Figure 3A). The contractile response to endothelin 1 from the pregnant thoracic aorta segments was increased in both (S)-MWCNTs and 10% surfactant exposed groups when compared to the naïve. There was a noticeable relaxation response to highest concentration of endothelin 1 in the (S)-MWCNT exposed pregnant group (Figure 3C). In contrast, the contractile responses to phenylephrine and endothelin 1 were not affected by (S)-MWCNT or 10% surfactant exposure in the non-pregnant female rats (Figure 3B and D).

The acetylcholine (0.001–10 μM) mediated relaxation response was not different in the pregnant group (Figure 3E), but was increased in both (S)-MWCNTs and 10% surfactant exposed non-pregnant aortic segments when compared to the naïve (Figure 3F). The EC$_{50}$ values were not different for the contractile and relaxation responses following (S)-MWCNT exposure (Supplementary Table 3).

**Responses of arterial segments 24 hours post-exposure to intravenous (IV) administration (D)-MWCNTs or DPPC/RSA in pregnant rats**

Twenty four hours following IV administration in pregnant rats, both (D)-MWCNTs and DPPC/RSA increased the maximal stress generation in the main uterine artery segments to a similar magnitude (3 – 4 mN/mm$^2$) when compared to naïve vessel segments with a similar concentration-response profile for the agonists: phenylephrine, endothelin 1 and angiotensin II (Figure 4A–C). The DPPC/RSA exposure elevated the baseline stress level of the uterine vessel segments while the (D)-MWCNTs exposure had no additional effect. The relaxation responses of the main uterine artery to acetylcholine were not changed by IV (D)-MWCNT or DPPC/RSA exposure (Figure 4D). We did not proceed to do non-pregnant comparisons in this exposure group as the differences in the contractile responses were attributed sole to DPPC/RSA suspension and not to MWCNT exposure.

An increase in contractile response in the mesenteric artery segments was seen at higher concentrations of phenylephrine following (D)-MWCNT exposure (supplementary Figure 2A). All other contractile/relaxation responses of the mesenteric artery and aortic segments were not significantly different between the (D)-MWCNTs or DPPC/RSA exposure groups (supplementary Figures 2B–D and 3A–C). Unlike in the uterine artery, DPPC/RSA did not
have a significant effect on the baseline stress level of the mesenteric artery or thoracic aorta. The EC\textsubscript{50} values for all responses are reported in supplementary Tables 1–3 and were not different with the exception for endothelin 1, where the EC\textsubscript{50} was reduced in the in the thoracic aortic segments following (D)-MWCNTs (13.3 ± 5.1 nM) or DPPC/RSA (12.4 ± 5.1 nM) exposure when compared to the naïve (71.9 ± 17.8 nM, Supplementary Table 3).

**Contribution of Rho kinase activity on the vascular tissue contractility following exposure to MWCNTs**

**Maintenance of stress in the presence of Rho kinase inhibitor**—Minor differences were observed in the relaxation responses to cumulative concentrations of the Rho kinase (ROCK) inhibitor, HA1077, during the stable phenylephrine pre-contraction for segments from all three vascular beds, regardless of the pregnancy state or route of exposure to the MWCNT (Figure 5 and Supplementary Figure 4). The EC\textsubscript{50} values for the concentration responses are reported in Supplementary Tables 1–3 and were not significantly different following MWCNT exposure except within the IV (D)-MWCNT exposure group during pregnancy (2.6 ± 0.2 μM compared to 1.8 ± 0.3 μM in the DPPC/RSA exposed group).

**RhoA, ROCK and eNOS mRNA and protein expression in rat aortic endothelial cells**—The mRNA expression of RhoA, ROCK1, ROCK2 and eNOS was not significantly changed in RAEC with 2–12 hour treatment with (S)-MWCNTs or (D)-MWCNTs when compared to untreated cells and vehicle controls (data not shown). Similarly, the protein expression of RhoA, ROCK and eNOS were not changed with 12 hours in vitro exposure to 10 μg/cm\textsuperscript{2} of (S)-MWCNTs or (D)-MWCNTs as assessed by the In-cell Western Assay (Supplementary Figures 5 and 6).

**Changes in the fetal components following MWCNT exposure**

**Changes in umbilical vein contractility**—The reactivity of the umbilical vein (vessel from the placenta to the fetus) was assessed following both IT and IV administration. Stress generation during K\textsuperscript{+}PSS and 1 μM of thromboxane mimetic (U46619) stimulations were not significantly different in umbilical vessel segments between MWCNT exposed (by either exposure route) and naïve animals (Figure 6A–D). The umbilical vein segments did not respond to acetylcholine and the relaxation response to 1μM sodium nitroprusside with a stable U46619 pre-contraction was not different following MWCNT exposure (Figure 6E and F).

**Changes in fetal and placental weight**—Mean weights of pregnant dams at the time of sacrifice were not significantly different between treatment groups (mean ± SEM): (S)-MWCNTs 298.2 ± 12.0 g (n=6), 10% surfactant 291.0 ± 10.8 g (n=6), (D)-MWCNTs 305.8 ± 7.8 g (n=6), DPPC/RSA 333.4 ± 24.9 g (n=6), and naïve 287.2 ± 10.9 g (n=10). The mean and range of litter size were also not different between the exposure groups: (S)-MWCNTs 10.7 (8–13), 10% surfactant 10.5 (10–11), (D)-MWCNTs 9.8 (9–11), DPPC/RSA 10.5 (9–12) and naïve 10.6 (8–13). Mean weights of the fetuses are reported in Figure 7A and B, after grouping them according to gestational day (GD). The mean fetal weight was reduced at GD 19 following MWCNT and dispersion media exposure by both routes and was evident.
across all gestational days studied following intravenous exposure. Gross external morphological abnormalities were not seen in the fetuses. An increase in the mean placental weight was observed following MWCNT exposure by both routes on GD 18 (Figure 7C and D).

**Fetal serum cytokine analysis**

The cytokines levels in the fetal serum were not significantly changed following exposure to MWCNT or dispersion media by either route of administration and are reported in Table 2.

**Discussion**

Twenty hours post exposure to MWCNT by either intratracheal or intravenous administration resulted in a limited alteration in isolated blood vessel segments’ responses to various pharmacological agents. The intratracheal instillation of 100 μg/kg of (S)-MWCNTs, was of note for we observed an increase in the contractile response to angiotensin II in the main uterine artery when compared to the response from the non-pregnant state. This increase in of stress generation of the uterine artery following MWCNT exposure was confined to the late gestational stage. Minimal changes in the contractile responses due to MWCNT exposure were seen in vessel segments from the other vascular beds studied in both pregnant and non-pregnant state. The enhanced contractile responses were not associated with comparable changes in relaxation responses with Rho kinase inhibition, suggesting that mechanisms other than RhoA/Rho kinase may underlie alterations in contractile responses. To our knowledge, this is one of the first attempts to identify pregnancy related changes in the contractile responses of several vascular tissues following exposure to MWCNTs by different routes.

The physiochemical characteristics of the suspensions of the MWCNTs are important in understating any delivery characteristics. According to Henderson et al, the pulmonary responses are similar in response to particle exposure by either inhalation or instillation, provided similar lung burdens [37]. Intratracheal instillation has been suggested to deliver less well dispersed MWCNTs to the lung epithelium resulting in fewer adverse effects when compared to short term inhalational exposure [34,38]. On the other hand, instillation exposes the animal to an acute, higher concentration of nanoparticles compared to inhalational exposure over a long period [34], bypassing the nasal cavity. Accounting for these conditions, the results from our IT exposure might be used to speculate on the outcomes of long term inhalational exposure. Exposure levels have been identified in laboratory and industrial facilities [15] handling MWCNTs. Current proposed guidelines by the National Institute for Occupational Safety and Health (NIOSH) limits nanotube exposure to 7 μg/m³ [39] and a human occupational exposure of 5 mg/m³ during a 8-hour day and 40-hour work week equate to approximately 20 μg of SWCNT aspiration in a mouse model [40]. A recent publication by Erdely et al. [41] identified a mean airborne mass concentration of 10.6 μg/m³ MWCNTs across eight different facilities that handle carbon nanotubes in the United States, which was calculated to correspond with a human alveolar deposition of 4.07 μg/day [41]. In a study designed to mimic such conditions while weighing and moving dry materials, MWCNT were found to range from 4,514–123,403 particles/L of air [42].

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Considering that average tidal volume in humans is 500 mL of air and average respiration rate is 12 breaths a minute (i.e. with a resting respiratory minute volume of 5 L/min), humans breathe approximately 360 L air/hour on average, this could result in a deposition of 1,625,040 – 44,425,080 MWCNT particles into the lungs in 1 hour. Based on calculations from flow-cytometry data (not shown), a 100 μg MWCNT exposure mass translated to approximately 743,000 particles per rat. Employing these most recent occupational exposure values (10.6 μg/m$^3$ [41]) it would require between 3–107 hours of constant occupational exposure to achieve a similar dosing used in our studies. Considering these conditions, and compared to other inhalational/instillation studies [13], 100 μg/kg dose used in the current study is a moderate - high dose. We choose to use an equivalent dose of MWCNTs in our IV exposure model to identify any effects of exposure to the same mass based dose of MWCNTs by two different routes. Future dose response studies including both low and high levels of exposure during pregnancy may be beneficial before applying these findings for regulatory purposes.

Previous studies with inhalational exposure to MWCNTs for 24–168 hours have identified a maximal impairment of endothelium dependent relaxation in coronary arterioles at 24 hours [11]. Therefore, the 24-hour post-exposure time point in this study should be conducive to identify changes in the contractile responses in extra-pulmonary vascular beds. Even during intravenous exposure, MWCNTs redistribute mainly to the lung and liver tissues [43] as pulmonary and hepatic circulations may be functioning as a filters during the first pass metabolism. The pulmonary responses to MWCNT deposition in such circumstances may be initiating the inflammatory cascade, contributing to changes in the contractile responses.

Several factors including the properties of MWCNT suspensions, route of exposure, pulmonary/systemic inflammatory response, pregnancy related physiological changes and sensitivity of the vascular bed may contribute to the differential contractile responses of vascular tissues that we have observed in this study. The reported zeta potentials of (S)-MWCNTs and (D)-MWCNTs suggest that both suspensions have a relatively good stability with minor agglomerate formation. The minor differences in the hydrodynamic size and zeta potential in different suspensions may not alone contribute significantly to modification of the vessel behavior via MWCNT exposure, as seen in different routes of exposure.

Previously reported distribution of MWCNTs following intravenous administration suggests that majority of the particles are distributed in the lungs following their first pass in circulation [23]. On the other hand, these particles are primarily distributed in the lungs following intratracheal instillation/pulmonary exposure [44]. Considering these distribution patterns by both routes, we chose to study the immune mediated pulmonary responses by analyzing the cell counts in a broncoalveolar lavage. We report a pregnancy related increase in the BAL cell counts suggesting an inflammatory response, reported as higher percentages of macrophages with both dispersion media and (D)-MWCNTs and the increased neutrophils with (S)-MWCNT exposure. Comparable changes in neutrophil and eosinophil counts were seen with pulmonary exposure to increasing doses of (S)-MWCNTs in a male mouse model [4]. These pulmonary responses following exposure to dispersion media or MWCNTs are also compatible with changes in the cytokine profiles, increased vascular tissue contractility and reduction in the fetal weight gain. The changes in the maternal serum
cytokine levels reported in our study does not indicate any preferential immune response as IFNγ (Th1), IL6 and IL10 (Th2) levels are not affected following exposure to MWCNTs by either route of administration. The increase in TNFα levels appears to be robustly influenced by both dispersant media (10% surfactant and DPPC/RSA) rather than MWCNTs and may potentially induce a Th1 type response, which can be detrimental during pregnancy (36). This conclusion is also supported by the reduction in fetal weight seen with both (D)-MWCNTs and DPPC/RSA. An increase in IL1β level was seen with both 10% surfactant and DPPC/RSA suggesting an immunological response to both dispersion media, which was down-regulated by the addition of MWCNTs. Previous studies have reported both Th1 and Th2 type immune responses following acute exposure to MWCNTs with increased levels of: TNFα, IL1β, IL6, IL10, and MCP1 (33, 38) early after exposure and reported to wane over time (39), supporting the low levels we measured at 24 hours post-exposure.

The overall stress generation in segments of the main uterine artery, from the naïve pregnant rats was lower compared to the response from naive non-pregnant rats, a response reflecting normal vasodilatory vascular behavior associated with pregnancy [18,19]. However, the uterine artery did present with an augmented response to Angiotensin II. Angiotensin II mediated responses are altered in the uterine vasculature during adverse pregnancy states such as pre-eclampsia where the AT1 receptor mediated constrictor function of angiotensin II predominates over the AT2 receptor mediated vasodilatory function [45,46]. The increased in contractility observed in the gestational uterine artery segments in our study could be mediated by alteration of either/both AT1 and AT2 receptor function following MWCNT exposure. Additionally, the calculated EC50 values for endothelin 1 were different in segments from all three vessels studied, suggesting an altered sensitivity to endothelin 1 or changes in endothelin receptor distribution with pregnancy. In contrast to the non-pregnant group, a significant increase in the contractile responses was evident only with the pregnant uterine artery segments, following (S)-MWCNT exposure. These observations suggest that pregnancy may render the uterine vasculature more susceptible to MWCNT exposure induced changes in contractility. Therefore, our observation of an increase in the contractile response or shifts in EC50 values following (S)-MWCNT exposure suggests there may be an adverse pregnancy outcome following pulmonary MWCNT exposure during pregnancy.

Compared to the uterine vasculature, the overall stress generation of mesenteric artery and thoracic aortic segments in response to contractile stimulation were not significantly different between naïve pregnant and non-pregnant animals. This relationship was not altered following MWCNT exposure by either route. The only difference in responses was observed at higher concentration of serotonin in the mesenteric artery, where we observed a higher stress generation/lower relaxation response following (S)-MWCNT exposure that could be mediated by alteration in the serotonin receptor profiles following MWCNT exposure. We interpret the contrast in response of the vessels from the different vascular beds to be related to the limited extent of remodeling that occurs in the mesenteric artery and thoracic aorta with pregnancy and renders them less vulnerable to the influence of MWCNT exposure.

The alterations in contractile responses reported in this study (in response to angiotensin II and serotonin) are similar to the potentiation of stress generation reported with other
cardiovascular pathologies, linked with elements of calcium sensitization and regulation of
the contractions by RhoA/Rho kinase pathway [47,48]. As reported in Figure 5 and
Supplementary Figure 4, there were only minor differences in the sensitivity to Rho kinase
inhibition in all three vascular beds following exposure to MWCNTs or dispersion media by
either route of exposure. Thus a response compatible with action of Rho kinase as being
responsible for the augmented contractile responses was not evident with MWCNT
exposure. Additionally, the lack of changes in Rho associated proteins and eNOS in rat
aortic endothelial cells exposed to MWCNTs in vitro would suggest that this pathway is not
mediating changes in the endothelial cell contribution to the augmented stress through
regulation of eNOS as reviewed by Yao et al. [25] and Satoh et al. [48]. Thus in aggregate
these data suggest that the rho kinase signaling was not a primary mechanism responsible
for augmented stress generation observed following MWCNT exposure. Other mechanisms
postulated to enhance force generation can include generation of reactive oxygen species
[49], increased oxidative stress [50] and enhanced cyclooxygenase signaling [51]. SWCNTs
have shown to increase oxidative stress and alter the mitochondrial signaling following
intrapharyngeal instillation [52]. MWCNTs may also affect the cardiovascular functions by
comparable mechanisms but have yet to be fully investigated.

Previous in vitro studies including proteomics analysis have shown that cellular functions
and pathways comprising generalized gene transcription and protein translation are affected
by direct exposure to MWCNTs [33,53]. Alternatively, the changes in the vascular system
may be mediated through bronchial epithelial cell release of pro-inflammatory cytokines
into circulation, such as IL6 and IL8 [6]. MWCNTs can also translocate following
pulmonary exposure via instillation and reach the extra-pulmonary sites [8,54], including the
vasculature leading to a local inflammatory response. Our previous in vitro studies done
with human aortic endothelial cells indicate increased expression of endothelial
inflammatory markers following exposure to (S)-MWCNTs [33] and may support a
mechanism for cytokine production which can influence vascular tissues.

When trying to understand the IT exposed MWCNT induced changes, it is important to
recognize that 10% surfactant used as a vehicle for suspending the MWCNTs also induces a
notable increase in stress generation in response to agonist stimulation compared the
constrictive responses from naïve animal group. We suggest that (S)-MWCNTs may have a
combined effect of both MWCNTs and surfactant and this effect is clearly demonstrated in
response to angiotensin II in the main uterine artery segments during pregnancy. However,
synthetic lung surfactant based suspensions are established for studying pulmonary exposure
effects of MWCNTs [55] and we chose to use the same for our study and were surprised to
see such a vascular response effect. The responses seen with intravenous exposure to
MWCNTs appear to be due to properties of the dispersant medium rather than due to
nanotubes as DPPC/RSA significantly increases the baseline stress generation. The different
media for the two routes of exposure were selected to simulate the biological media that area
related to the exposure routes and were speculated to have no/minimal effects on vascular
contractile responses. The dispersant medium is known to affect the cellular uptake of the
nanoparticles [56] and presumed to impact overall cellular function. The dispersant media
are known to contribute to the composition of protein or lipid corona associated with the

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nanoparticles in the biological systems [57,58]. It may be likely that the corona on these MWCNT is different enough to mask a significant MWCNT effect.

Neither MWCNTs nor dispersion medium induced significant changes in contractile responses of umbilical vein segments, suggesting that these exposures may only be affecting the maternal side of the circulation. However, detrimental effects were seen in the mean fetal weight following MWCNT exposure via both routes along with a significant contribution by the dispersion media alone. The IV exposure to (D)-MWCNTs appeared to be more effective at reducing the fetal growth earlier in gestational exposure window studied, whereas the weight reduction following IT exposure is mainly attributed to dispersion media. The placental transfer of the nanoparticles is affected by multiple factors including the particle size, dispersion medium, and the stage of the pregnancy [59] which could contribute to effects on fetal growth. Additionally, fetal microvessel dysfunction following exposure to engineered nanomaterials which was recently proposed by Stapleton et al. [11] may be a possible underlying explanation for our observations of reduced fetal weight despite the absence of augmented contractile responses in umbilical circulation. Stapleton et al. [11] used the fetal tail artery as a representative vessel from the fetal microcirculation and reported decreased responses in both endothelium dependent and independent relaxation [60]. Their findings suggests the applicability of the Barker Hypothesis (i.e. the relation between retarded growth in early life and risk of adult disease is due to long term effects on physiology and metabolism imposed by an adverse environment during critical periods of development) to explain the changes observed in the fetus following maternal nanoparticle exposure [60,61]. This hypothesis may also hold true for our MWCNT exposure scenario, suggesting that the differences in the fetal weight gain may be a reflection of limited blood supply due to increased contraction observed in the uterine vascular segments.

**Conclusions**

In conclusion, the observations in this study suggest that vascular contractility may change following MWCNT exposure depending on multiple factors, including life stage (pregnant or non-pregnant), route of exposure, MWCNT dispersion media and the target vascular bed. Multiple agonist-mediated responses are differentially affected between the pregnant and non-pregnant stages with a significant increase in the stress generation of the uterine artery in response to angiotensin II confined to the pregnant stage. These agonists alter the contractile mechanism through various signaling cascades and we assessed the contribution RhoA/Rho kinase pathway in mediating these responses. We were unable to demonstrate that the RhoA/Rho kinase signaling cascade was significantly altered in response to MWCNT exposure and could not account for the augmented contractile responses, suggesting that other pro-constrictor mechanisms are likely to be involved. Our comparisons with naïve rats revealed a significant influence of the dispersion media on vascular tissue contractility and fetal weight gain, suggesting MWCNT exposure in isolation has no/ minimal effects under the exposure scenarios applied in this study.

**Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.
Acknowledgments

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Abbreviations

<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>5HT</td>
<td>Serotonin</td>
</tr>
<tr>
<td>Ach</td>
<td>Acetylcholine</td>
</tr>
<tr>
<td>ANG II</td>
<td>Angiotensin II</td>
</tr>
<tr>
<td>ANOVA</td>
<td>Analysis of Variance</td>
</tr>
<tr>
<td>BAL</td>
<td>Bronchoalveolar Lavage</td>
</tr>
<tr>
<td>(D)-MWCNTs</td>
<td>MWCNT suspended in DPPC and RSA based medium</td>
</tr>
<tr>
<td>DPPC1</td>
<td>2-dipalmityl-sn-glycero-3-phosphocholine</td>
</tr>
<tr>
<td>DPPC/RSA DPPC</td>
<td>RSA and Phosphate Buffered Saline-based Medium</td>
</tr>
<tr>
<td>GD</td>
<td>Gestational Day</td>
</tr>
<tr>
<td>EC50</td>
<td>Half-maximal Effective Concentration</td>
</tr>
<tr>
<td>Enos</td>
<td>Endothelial Nitric Oxide Synthase</td>
</tr>
<tr>
<td>IL1β</td>
<td>Interleukin 1 Beta</td>
</tr>
<tr>
<td>IL6</td>
<td>Interleukin 6</td>
</tr>
<tr>
<td>IL10</td>
<td>Interleukin 10</td>
</tr>
<tr>
<td>IFNγ</td>
<td>Interferon, gamma</td>
</tr>
<tr>
<td>IT</td>
<td>Intratracheal Instillation</td>
</tr>
<tr>
<td>IV</td>
<td>Intravenous administration</td>
</tr>
<tr>
<td>MCP1</td>
<td>Monocyte Chemotactic Protein-1</td>
</tr>
<tr>
<td>MWCNT</td>
<td>Multi-walled Carbon Nanotube</td>
</tr>
<tr>
<td>NP</td>
<td>Non-pregnant</td>
</tr>
<tr>
<td>P</td>
<td>Pregnant</td>
</tr>
<tr>
<td>PAI1</td>
<td>Plasminogen Activator Inhibitor-1</td>
</tr>
<tr>
<td>PE</td>
<td>Phenylephrine</td>
</tr>
<tr>
<td>PSS</td>
<td>Physiological Saline Solution</td>
</tr>
<tr>
<td>RAEC</td>
<td>Rat Aortic Endothelial Cells</td>
</tr>
<tr>
<td>(S)-MWCNTs</td>
<td>Multi-walled Carbon Nanotubes dispersed in 10% surfactant</td>
</tr>
</tbody>
</table>
TNFα  Tumor Necrosis Factor, alpha
VEGF  Vascular Endothelial Growth Factor

References


Figure 1. Changes in the contractile responses of the main uterine artery following intratracheal instillation (IT) of MWCNTs

The changes in the contractile responses as assessed by wire myography of the main uterine artery from 17 – 19 days pregnant (A, C, E and G) and non-pregnant female (B, D, F and H) Sprague Dawley rats, 24 hours following intratracheal instillation of 100 μg/kg of (S)-MWCNTs or 10% surfactant. The stress generation in response to cumulative concentrations of phenylephrine (PE; A and B), angiotensin II (ANG II; C and D) and endothelin 1 (ET-1; E and F) are plotted. The percentage relaxation from a 30 μM phenylephrine pre-stimulation...
stress level in response to cumulative concentrations of acetylcholine (Ach; G and H) is graphed. * indicates $p < 0.05$ compared to 10% surfactant while # indicates $p < 0.05$ compared to naïve using repeated measures ANOVA ($n = 5 – 7$). The $p$ values were derived following the comparison of each concentration response curve across treatment groups using a regression analysis by examining the best-fit values.
Figure 2. Changes in the contractile responses of the mesenteric artery following intratracheal instillation (IT) of MWCNTs

The changes in the contractile responses were assessed by wire myography of the first order mesenteric artery from 17 – 19 days pregnant (A, C, E and G) and non-pregnant female (B, D, F and H) Sprague Dawley rats, 24 hours following intratracheal instillation (IT) of 100 μg/kg of (S)-MWCNTs or 10% surfactant. The stress generation in response to cumulative concentrations of phenylephrine (PE; A and B), endothelin 1 (ET-1; C and D) and serotonin (5HT; E and F) are plotted. The percentage relaxation from a 30 μM phenylephrine pre-stimulation stress level in response to cumulative concentrations of acetylcholine (Ach; G and H) is graphed. * indicates p < 0.05 compared to 10% surfactant while # indicates p < 0.05 compared to naïve using repeated measures ANOVA (n = 4 – 7). The p values were
derived following the comparison of each concentration response curve across treatment groups using a regression analysis by examining the best-fit values.
Figure 3. Changes in the contractile responses of the thoracic aorta following intratracheal instillation (IT) of exposure to MWCNTs
The changes in the contractile responses were assessed by wire myography of the thoracic aorta from 17 – 19 days pregnant (A, C and E) and non-pregnant female (B, D and F) Sprague Dawley rats, 24 hours following intratracheal instillation of 100 μg/kg of (S)-MWCNTs or 10% surfactant. The stress generation in response to cumulative concentrations of phenylephrine (PE; A and B) and endothelin 1 (ET-1; C and D) are plotted. The percentage relaxation form a 10 μM phenylephrine pre-stimulation stress level in response to cumulative concentrations of acetylcholine (Ach; E and F) is graphed. * indicates p < 0.05 compared to 10% surfactant while # indicates p < 0.05 compared to naïve using repeated measures ANOVA (n = 4 – 8). The p values were derived following the comparison of each concentration response curve across treatment groups using a regression analysis by examining the best-fit values.
Figure 4. Changes in the contractile responses of the main uterine artery following intravenous administration (IV) of MWCNTs

The changes in the contractile responses were assessed by wire myography of the main uterine artery segments from 17 – 19 days pregnant (A, B, C and D) Sprague Dawley rats, 24 hours following intravenous administration of 100 μg/kg of (D)-MWCNTs or DPPC/ RSA. The stress generation in response to cumulative concentrations of phenylephrine (PE; A), angiotensin II (ANG II; B) and endothelin 1 (ET-1; C) are plotted. The percentage relaxation from a 30 μM phenylephrine pre-stimulation stress level in response to cumulative concentrations of acetylcholine is graphed (Ach: D). # indicates p < 0.05 compared to naïve.
using repeated measures ANOVA ($n = 5 – 8$). The $p$ values were derived following the comparison of each concentration response curve across treatment groups using a regression analysis by examining the best-fit values.
Figure 5. Changes in stress generation in the presence of a Rho kinase inhibitor following intratracheal instillation (IT) MWCNTs

The reduction in stress generation is reported as the percentage relaxation from a phenylephrine (30 μM for uterine/mesenteric arteries and 10 μM for aorta) pre-stimulation stress level in response to cumulative additions of a Rho kinase inhibitor (HA1077). All responses were assessed by wire myography, 24 hours following intratracheal instillation of 100 μg/kg of (S)-MWCNTs or 10% surfactant from 17 – 19 days pregnant (A, C and E) and non-pregnant female (B, D and F) Sprague Dawley rats. Panels A and B: main uterine.
artery; C and D: first order mesenteric artery; E and F: thoracic aorta. # indicates \( p < 0.05 \) compared to naïve using repeated measures ANOVA (n = 4 – 6). The \( p \) values were derived following the comparison of each concentration response curve across treatment groups using a regression analysis by examining the best-fit values.
Figure 6. Changes in contractile responses of the umbilical vein following maternal exposure to MWCNTs

The changes in stress generation in the umbilical vein segments were assessed by wire myography in response to 109 mM K$^+$ depolarization (A and B) and 1 μM thromboxane agonist (U46619, B and D) 24 hours post-exposure to intratracheal instillation (IT) of 100 μg/kg of (S)-MWCNTs or 10% surfactant (A and C) or intravenous administration (IV) of (D)-MWCNTs or DPPC/RSA (B and D), from 17 – 19 days pregnant Sprague Dawley rats ($n = 12$). The percentage relaxation in response to sodium nitroprusside (SNP) following U46619 (1 μM) pre-contraction in the umbilical vein 24 hours post-exposure to intratracheal...
instillation of (S)-MWCNTs or 10% surfactant (E) or intravenous administration of (D)-MWCNT or DPPC/RSA is graphed (F) ($n = 12$).
Figure 7. Changes in the fetal and placental weight following exposure to MWCNTs

Changes in fetal (A and B) and placental (C and D) weight 24 hours post-exposure to intratracheal instillation (IT) of 100 μg/kg of (S)-MWCNTs or 10% surfactant (A and C) or intravenous administration (IV) of (D)-MWCNTs or DPPC/RSA (B and D) in 17 – 19 days pregnant Sprague Dawley rats. Three pups/placentae were weighed from each dam included in the study and the number of pups/placentae are indicated within each bar. * indicates $p < 0.05$ when compared using one way ANOVA and Turkey post-hoc test. GD = day of gestation.
Table 1

Cytokine levels in maternal serum 24 hours post-exposure to MWCNTs

The maternal serum cytokines were evaluated using Milliplex MAP Cytokine/Chemokine Panel and Immunoassay (EMD Millipore MA, USA). The assays were run using Luminex 100/200 (Luminex, Austin, TX) and results reported using Luminex xPONENT® software versions 2.3/3.1. The mean and the SEM are reported for serum cytokines of pregnant and non-pregnant female Sprague Dawley rats (n = 5 – 8).

<table>
<thead>
<tr>
<th>Cytokine</th>
<th>NP-naive</th>
<th>NP-IT 10% surfactant</th>
<th>NP-IT (S)-MWCNTs</th>
<th>P-naive</th>
<th>P-IT 10% surfactant</th>
<th>P-IT (S)-MWCNTs</th>
<th>P-IV DPPC/RSA</th>
<th>P-IV (D)-MWCNTs</th>
</tr>
</thead>
<tbody>
<tr>
<td>IL1β (pg/ml)</td>
<td>56.0 ± 38.5</td>
<td>10.1 ± 3.8</td>
<td>7.3 ± 5.4</td>
<td>14.6 ± 7.1†</td>
<td>42.1 ± 7.1</td>
<td>15.8 ± 5.7°</td>
<td>70.2 ± 17.3#</td>
<td>45.0 ± 12.5</td>
</tr>
<tr>
<td>IL6 (pg/ml)</td>
<td>1245.0 ± 826.0</td>
<td>640.0 ± 208.2</td>
<td>485.0 ± 209.0</td>
<td>77.9 ± 72.8</td>
<td>215.3 ± 62.6</td>
<td>183.0 ± 128.8</td>
<td>126.8 ± 92.1</td>
<td>229.4 ± 115.5</td>
</tr>
<tr>
<td>IL10 (pg/ml)</td>
<td>21.7 ± 9.0</td>
<td>13.4 ± 4.8</td>
<td>10.2 ± 6.4</td>
<td>5.8 ± 3.7</td>
<td>9.6 ± 4.4</td>
<td>7.3 ± 3.6</td>
<td>16.4 ± 6.0</td>
<td>9.8 ± 4.2</td>
</tr>
<tr>
<td>INFγ (pg/ml)</td>
<td>338.4 ± 135.3</td>
<td>189.6 ± 33.6</td>
<td>174.1 ± 57.3</td>
<td>198.5 ± 50.5</td>
<td>106.4 ± 10.8</td>
<td>131.0 ± 36.8</td>
<td>273.1 ± 102.2</td>
<td>242.0 ± 50.1</td>
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<tr>
<td>MCP1 (pg/ml)</td>
<td>823.2 ± 223.9</td>
<td>467.0 ± 106.6</td>
<td>540.2 ± 248.9</td>
<td>288.1 ± 96.0</td>
<td>492.9 ± 27.0</td>
<td>336.2 ± 96.5</td>
<td>510.6 ± 54.8</td>
<td>478.2 ± 36.2</td>
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<tr>
<td>VEGF (pg/ml)</td>
<td>53.7 ± 15.4</td>
<td>30.1 ± 4.3</td>
<td>29.4 ± 7.5</td>
<td>510.2 ± 111.9</td>
<td>377.4 ± 81.1</td>
<td>476.6 ± 42.2</td>
<td>421.8 ± 63.0</td>
<td>432.8 ± 56.6</td>
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<tr>
<td>TNFa (pg/ml)</td>
<td>40.9 ± 12.83</td>
<td>30.3 ± 4.6</td>
<td>27.0 ± 8.9</td>
<td>6.8 ± 3.9†</td>
<td>45.3 ± 10.2</td>
<td>23.9 ± 9.9</td>
<td>43.7 ± 13.3#</td>
<td>34.1 ± 10.4#</td>
</tr>
</tbody>
</table>

IT: intratracheal instillation and IV: intravenous administration, P: pregnant and NP: non-pregnant, N/A: not available MWCNT: Multi-wall carbon nanotube, 10% surfactant: 10 % surfactant in saline, (S)-MWCNTs: MWCNT suspended in 10% surfactant, DPPC/RSA: vehicle used for IV MWCNT delivery and (D)-MWCNTs: MWCNT suspended in DPPC/RSA.

* indicates p < 0.05 when compared to the dispersion medium of each route,

# indicates p < 0.05 when compared to naïve and

† indicates p < 0.05 when compared to same treatment in non – pregnant.
Table 2

Cytokine levels in fetal serum 24 hours post-exposure to MWCNTs

The fetal serum cytokines (a pooled sample from three fetuses/dam) were evaluated using Milliplex MAP Cytokine/Chemokine Panel and Immunoassay (EMD Millipore MA, USA). These assays were run using Luminex 100/200 (Luminex, Austin, TX) and results reported using Luminex xPONENT® software versions 2.3/3.1. The mean and the SEM are reported for serum cytokines from fetuses from 17–19 days pregnant Sprague Dawley rats (n = 4–6).

<table>
<thead>
<tr>
<th>Cytokine</th>
<th>naïve (pg/ml)</th>
<th>IT 10% surfactant</th>
<th>IT (S)-MWCNTs</th>
<th>IV DPPC/RSA</th>
<th>IV (D)-MWCNTs</th>
</tr>
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<tbody>
<tr>
<td>IL1β</td>
<td>1455.0 ± 372.5</td>
<td>1597.0 ± 310.3</td>
<td>1657.0 ± 699.4</td>
<td>1932.0 ± 861.7</td>
<td>1500.0 ± 375.9</td>
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<tr>
<td>IL6</td>
<td>75.7 ± 16.8</td>
<td>277.0 ± 78.5#</td>
<td>64.9 ± 55.4</td>
<td>120.4 ± 86.5</td>
<td>58.7 ± 55.9</td>
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<tr>
<td>IL10</td>
<td>70.0 ± 17.0</td>
<td>76.0 ± 13.5</td>
<td>49.7 ± 6.1</td>
<td>117.2 ± 56.9</td>
<td>66.8 ± 12.2</td>
</tr>
<tr>
<td>INFγ</td>
<td>283.3 ± 42.5</td>
<td>324.5 ± 96.4</td>
<td>278.7 ± 30.6</td>
<td>405.8 ± 139.0</td>
<td>355.9 ± 139.0</td>
</tr>
<tr>
<td>MCP1</td>
<td>903.4 ± 310.8</td>
<td>986.7 ± 137.7</td>
<td>836.6 ± 60.3</td>
<td>991.2 ± 184.6</td>
<td>813.2 ± 113.3</td>
</tr>
<tr>
<td>VEGF</td>
<td>409.5 ± 83.0</td>
<td>483.8 ± 59.3</td>
<td>457.2 ± 52.2</td>
<td>620.2 ± 127.5</td>
<td>512.1 ± 47.0</td>
</tr>
<tr>
<td>TNFα</td>
<td>5.9 ± 5.9</td>
<td>3.3 ± 3.3</td>
<td>11.5 ± 5.9</td>
<td>6.7 ± 4.2</td>
<td>3.1 ± 2.3</td>
</tr>
</tbody>
</table>

IT: intratracheal instillation and IV: intravenous administration, MWCNT: Multi-wall carbon nanotube, 10% surfactant: 10% surfactant in saline, (S)-MWCNTs: MWCNT suspended in 10% surfactant, DPPC/RSA: vehicle used for IV MWCNT delivery and (D)-MWCNTs: MWCNT suspended in DPPC/RSA.

# indicates p < 0.05 when compared to naïve.